

Next Generation Sequencing

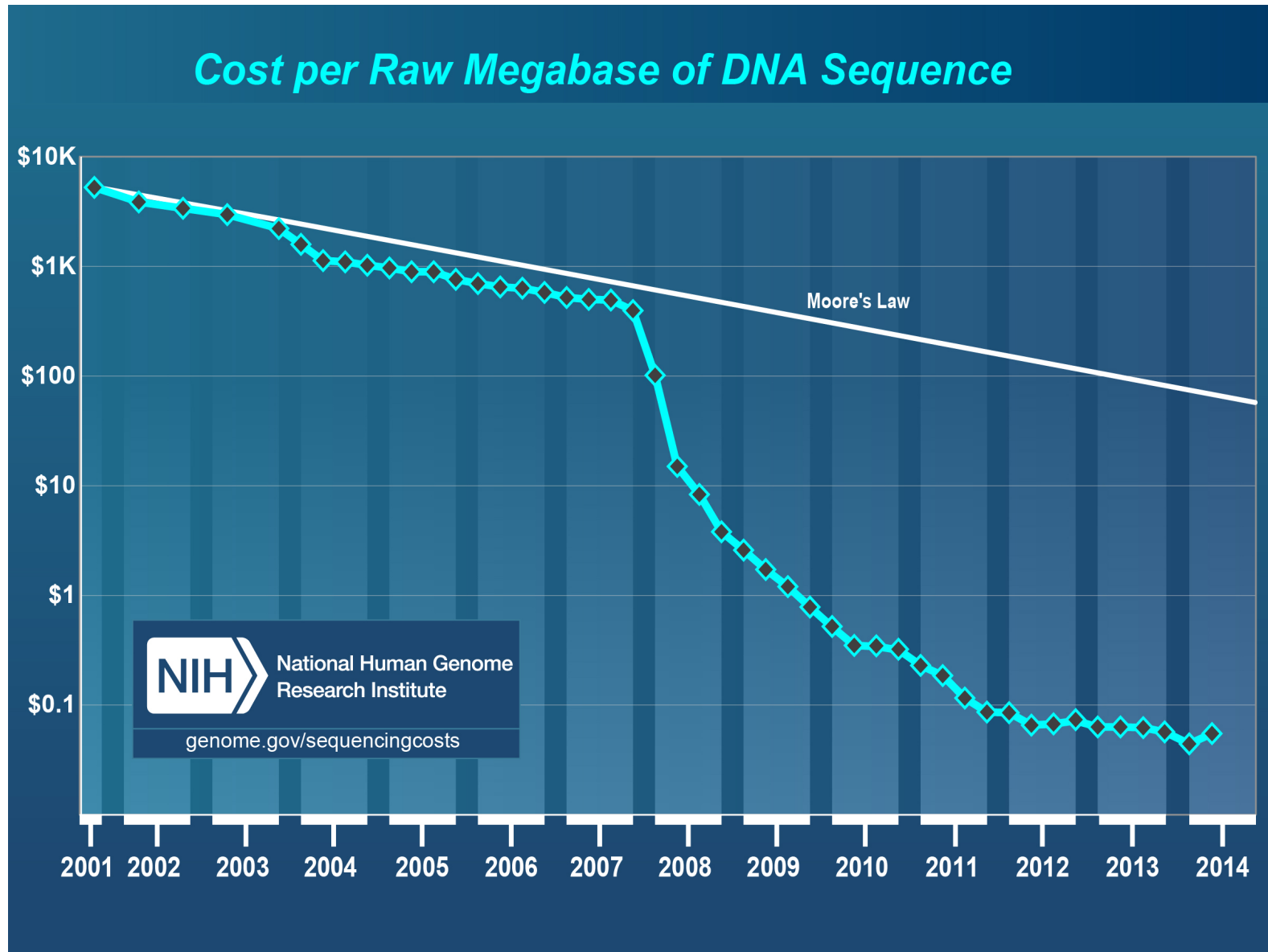
October 8, 2014

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Sequencing costs have dropped dramatically



Talk outline

- Traditional sequencing
- Next-generation sequencing
- Illumina sequencing
- NGS applications

Traditional Sequencing

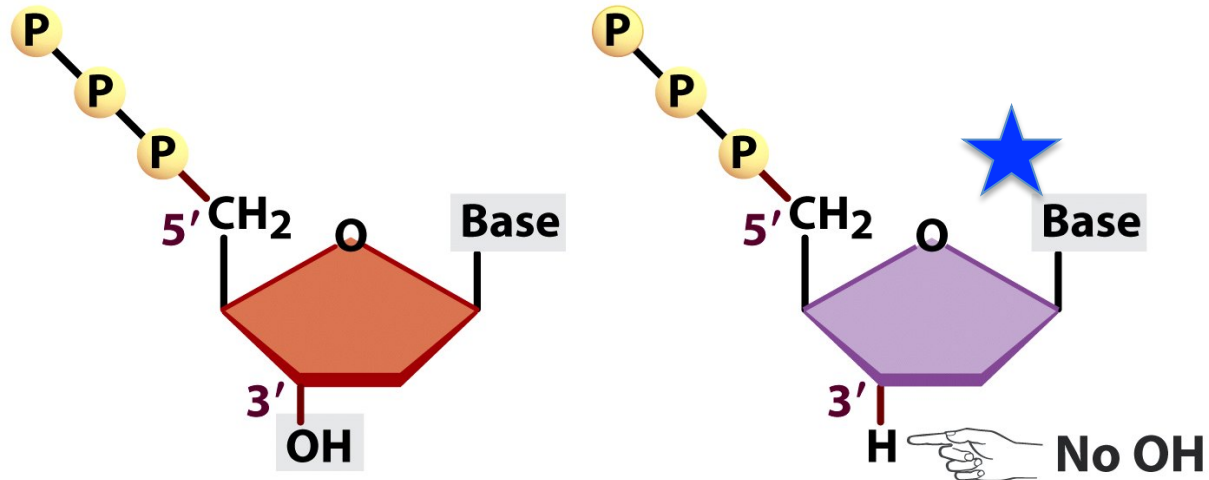
- Primer extension with labeled terminators
- 700 base read length

ACTAGCTGGACTCGTCACACT

Traditional Sequencing

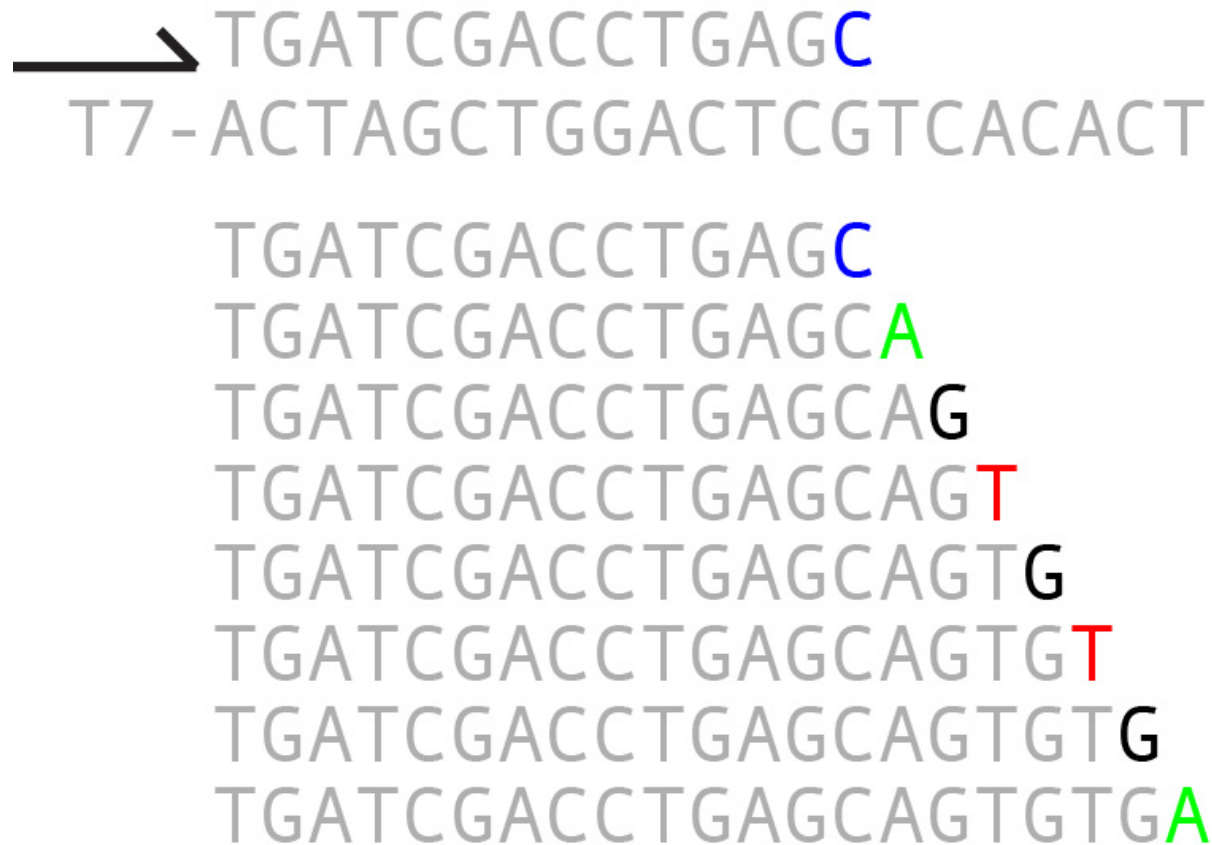
- Primer extension with labeled terminators
- 700 base read length

→ TGATCGACCTGAGC
T7 - ACTAGCTGGACTCGTCACACT



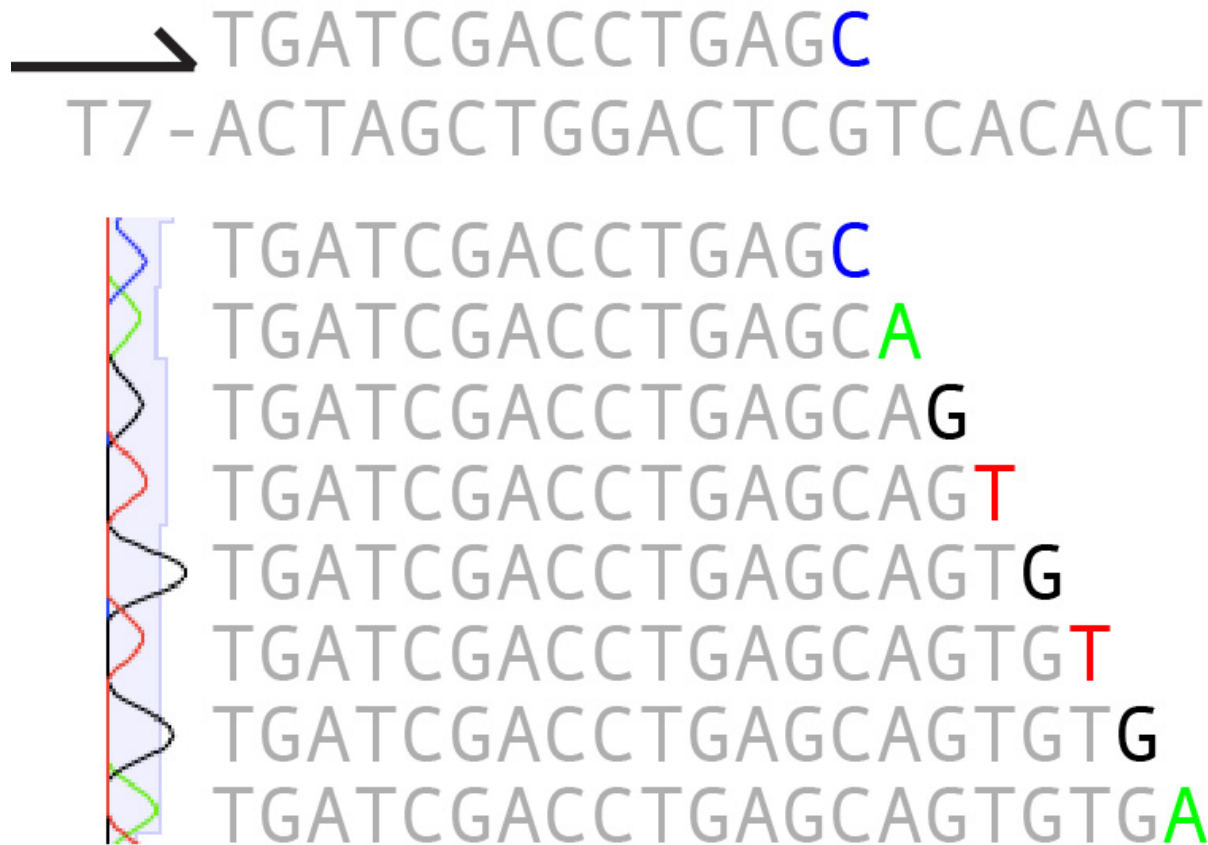
Traditional Sequencing

- Primer extension with labeled terminators
- 700 base read length



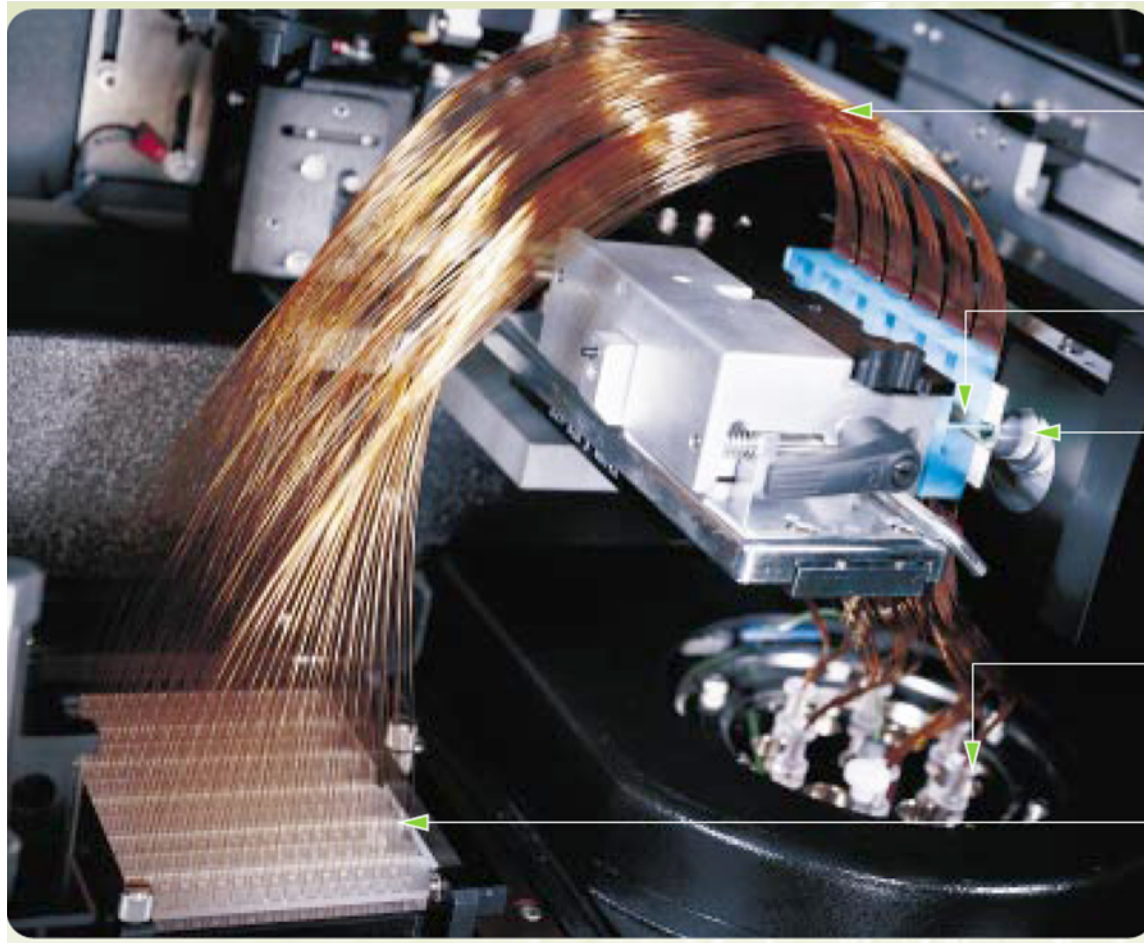
Traditional Sequencing

- Primer extension with labeled terminators
- 700 base read length



Traditional Sequencing

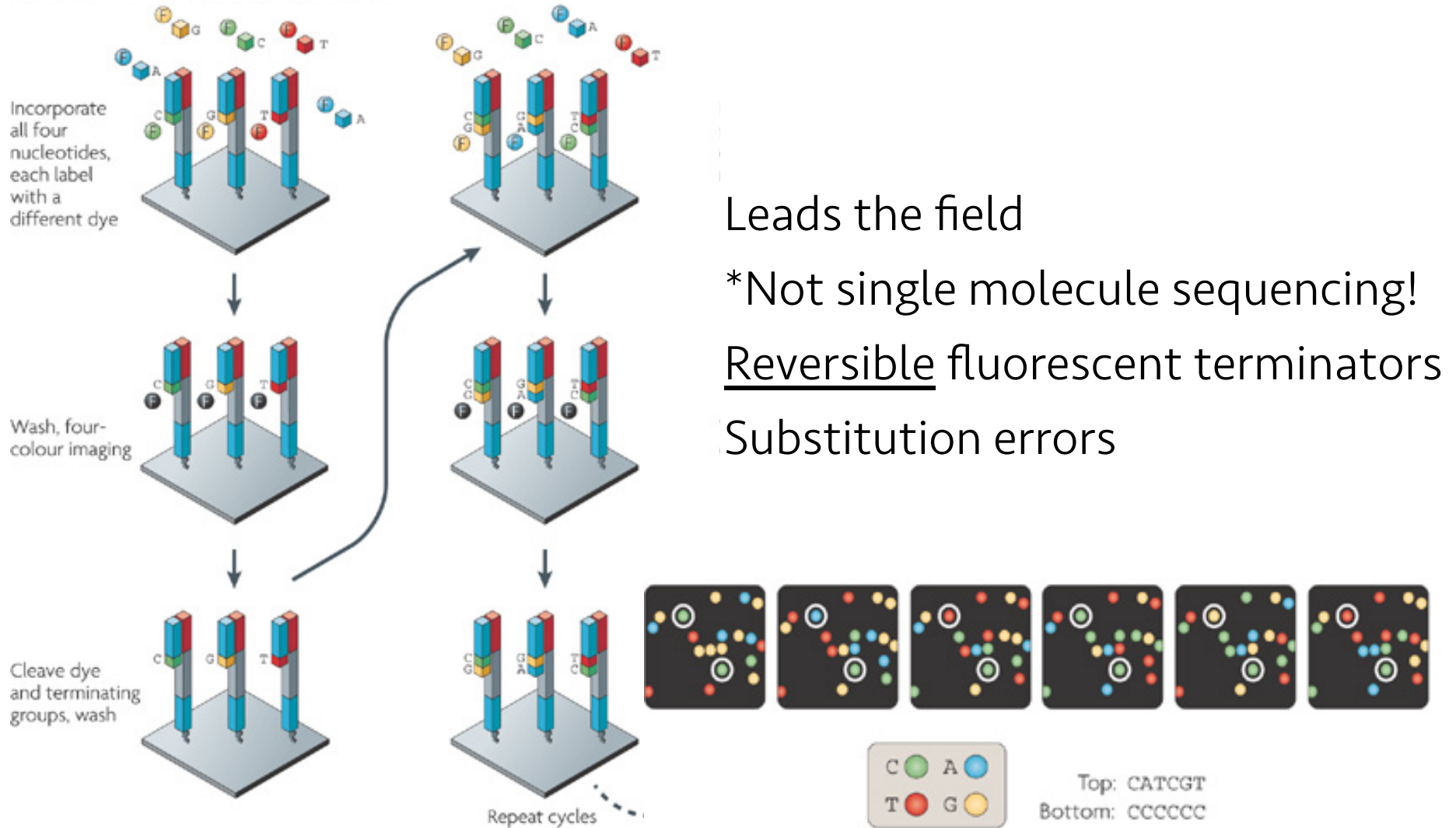
- Primer extension with labeled terminators
- 700 base read length



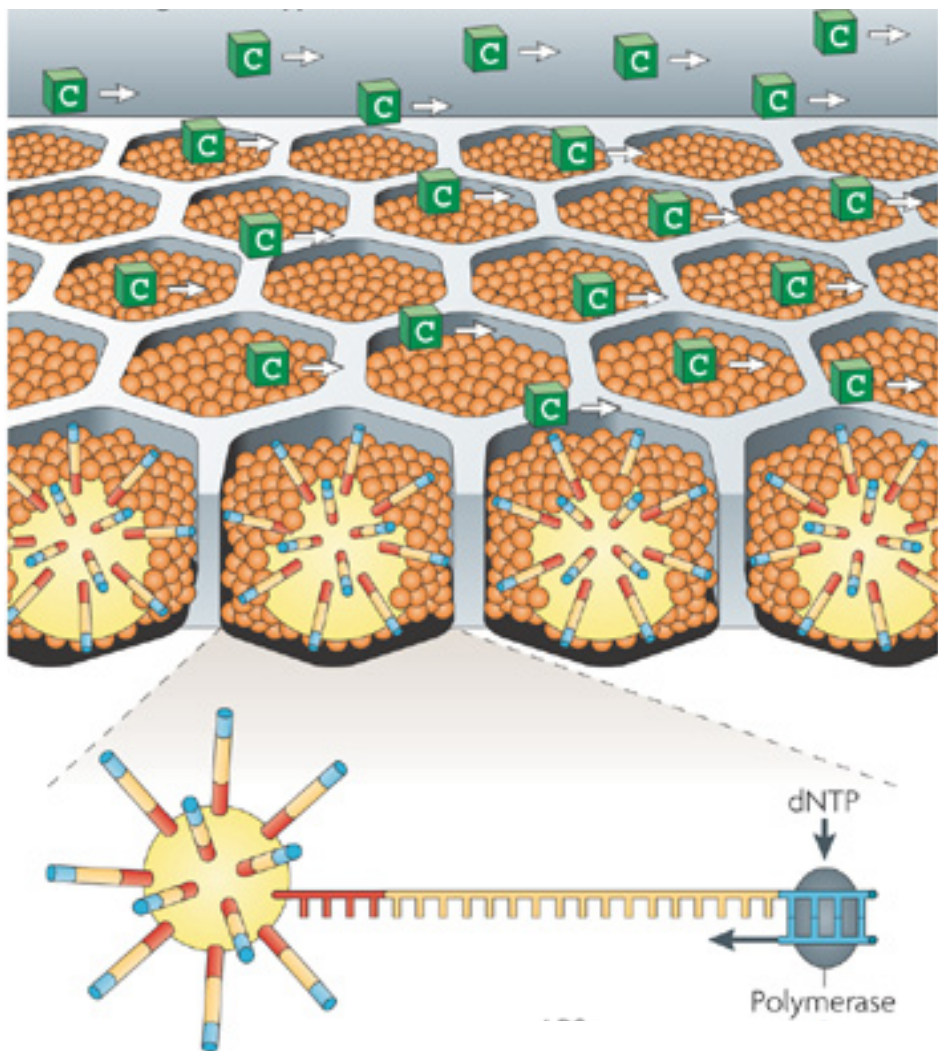
Next generation Massively Parallel Sequencing Technologies

- Many reads
- Next generation systems
 - Short read
 - Illumina – Sequencing by synthesis (dye)
 - Ion Torrent – sequencing by synthesis (pH)
 - Long read
 - Pacific Bioscience – single molecule (dye)
 - Oxford Nanopore – single molecule (current)

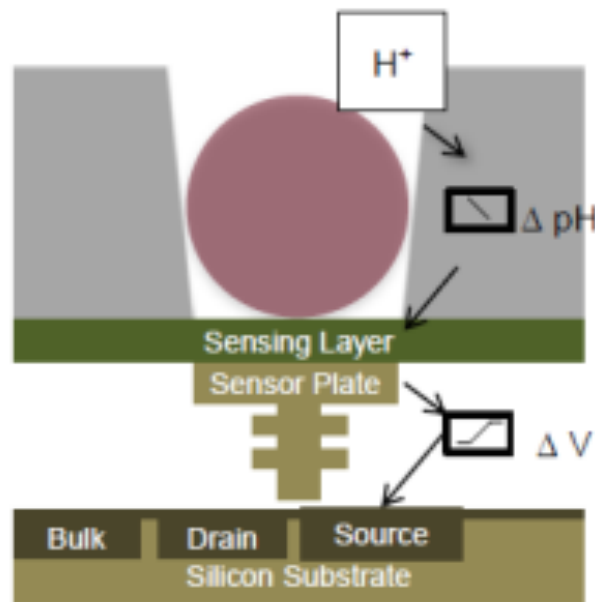
Illumina sequencing



Ion Torrent



Nature Rev Gen



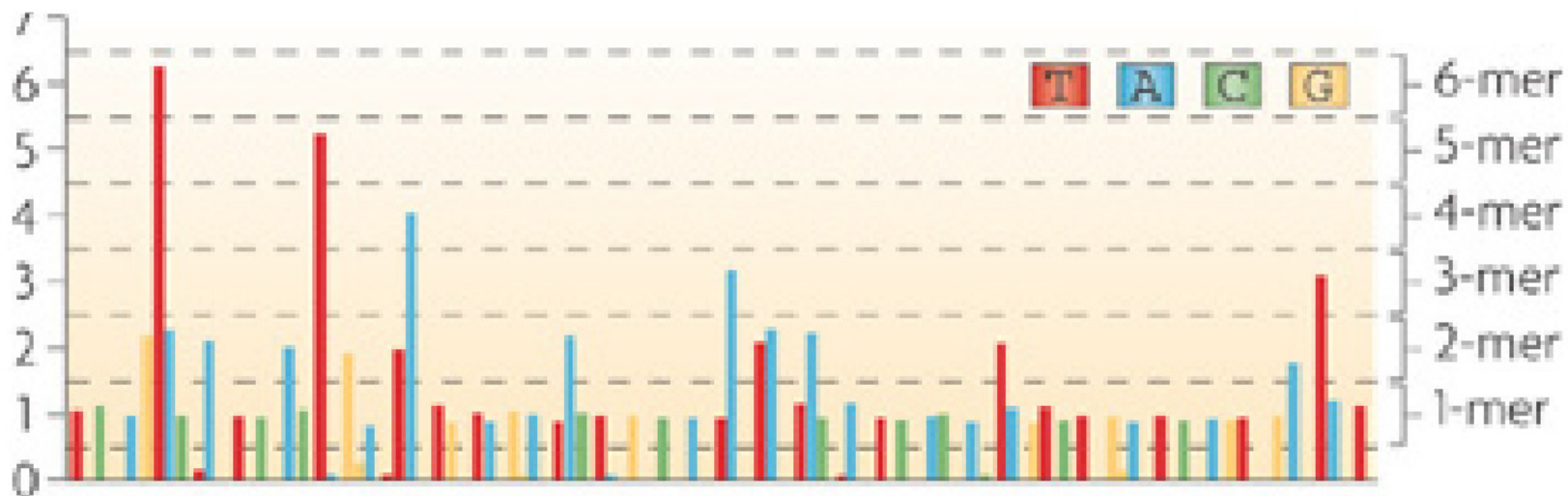
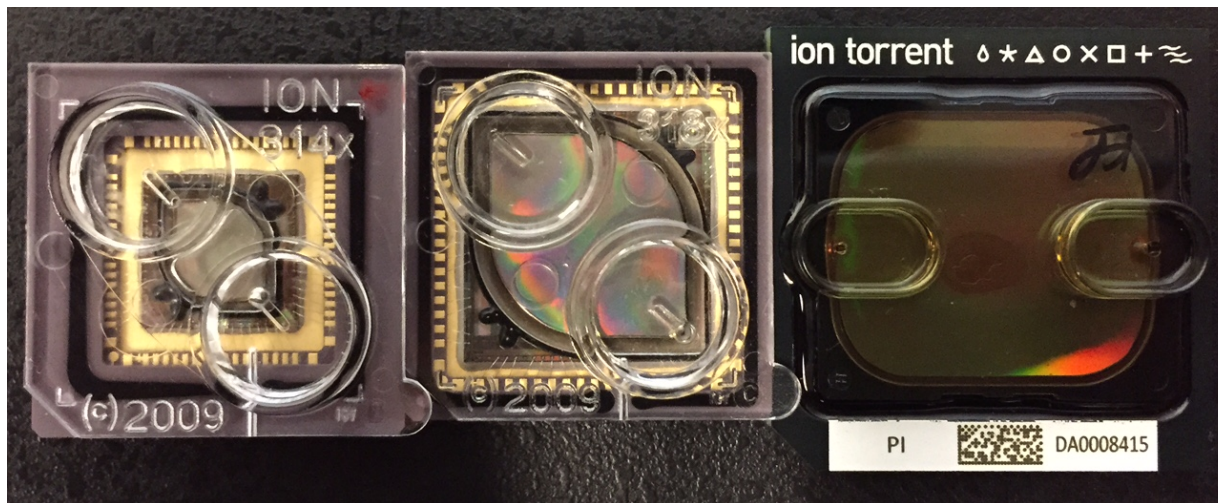
PGM

314 0.6M
316 3M
318 5.5M

Proton

PI 82M
PII 330M
PIII 660M

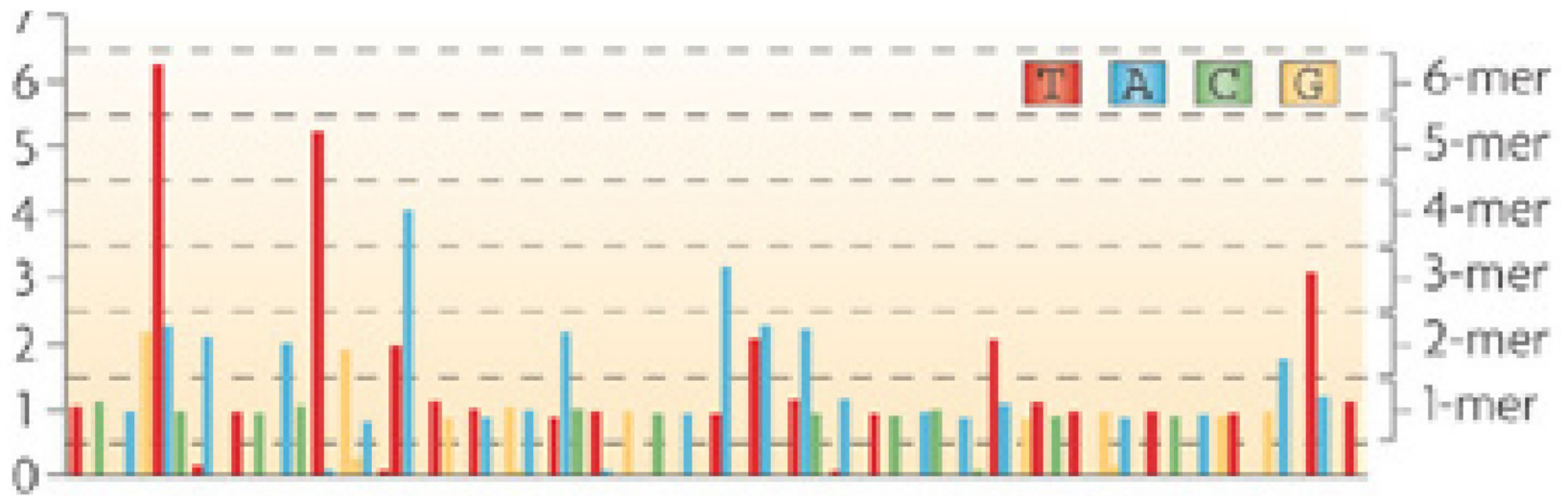
Ion Torrent



Nature Rev Gen

Ion Torrent

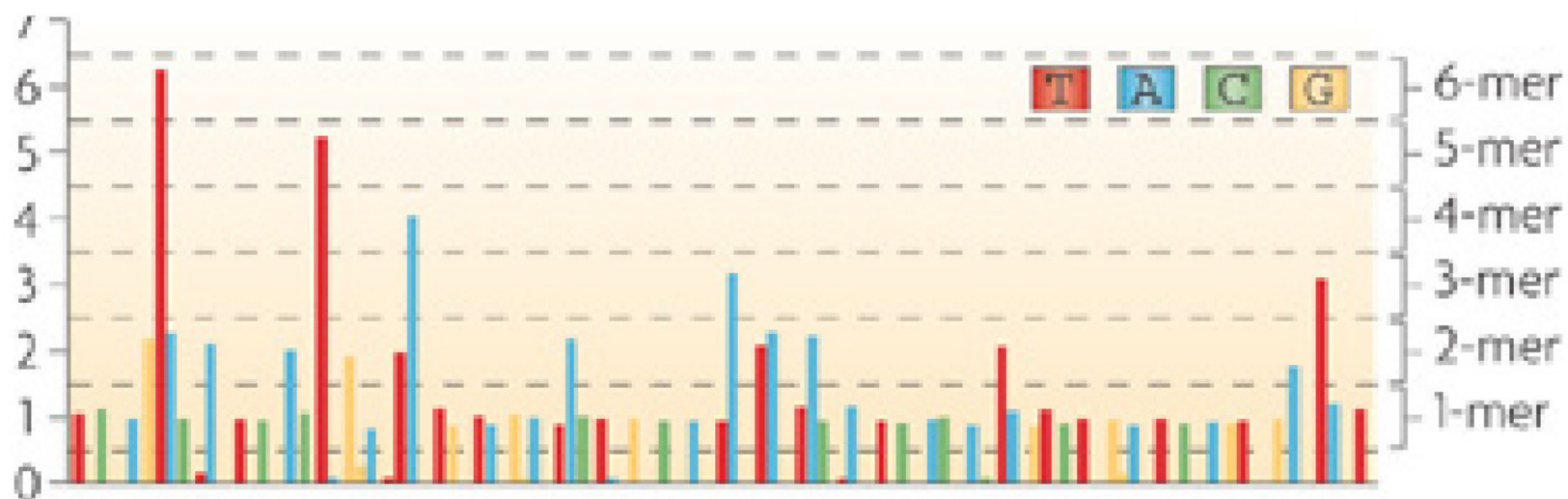
TCAGTACATCACTGATATAT
11126212112152124111



Ion Torrent

TCAGGTTTTTTAACAAATCAACTTTTTGGATTAAAATGT

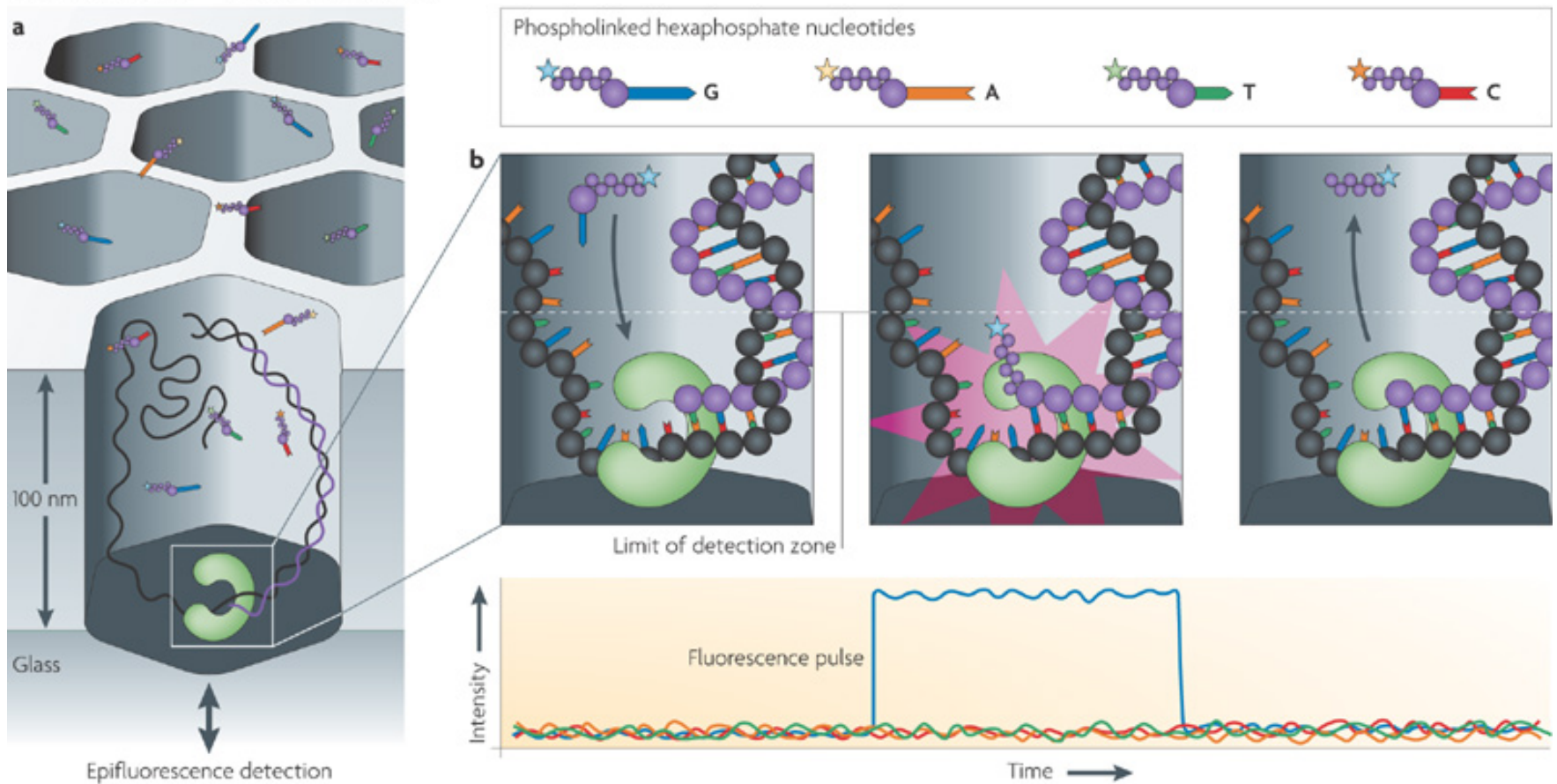
TCAGTACATCACTGATATAT
11126212112152124111



Insertion/deletion errors

Problems with scaling up

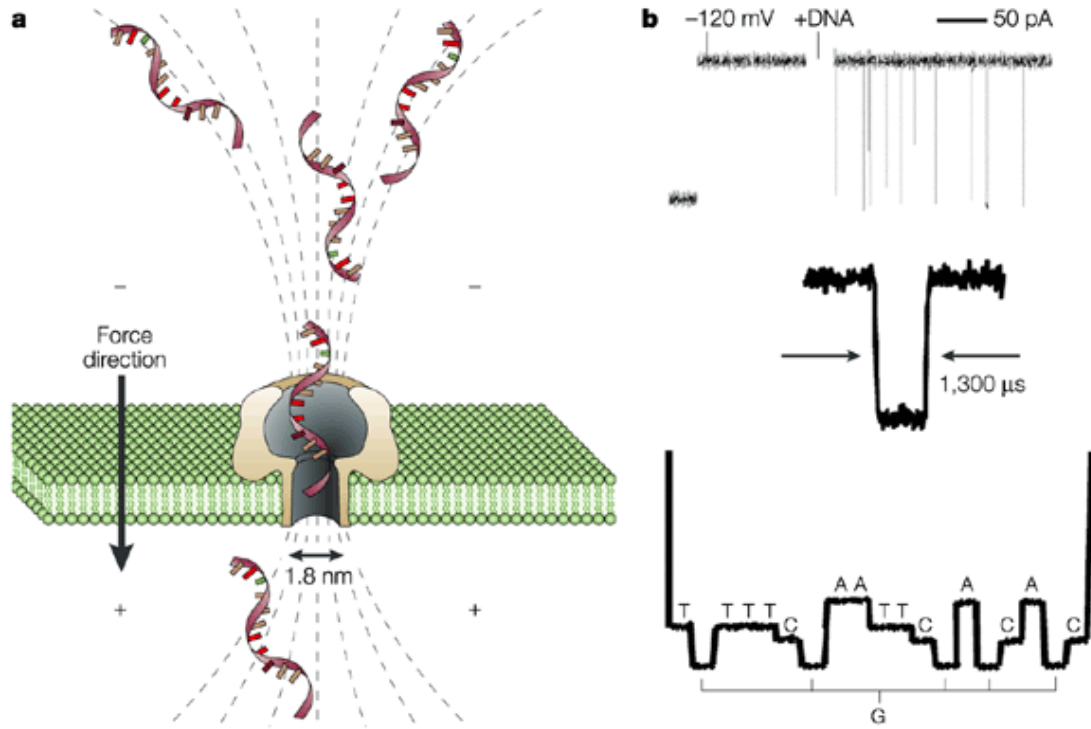
Pacific Biosciences



Nature Reviews | Genetics

Single molecule sequencing
20 kB read lengths
15% error rate

Oxford Nanopore



Nature Reviews | Drug Discovery

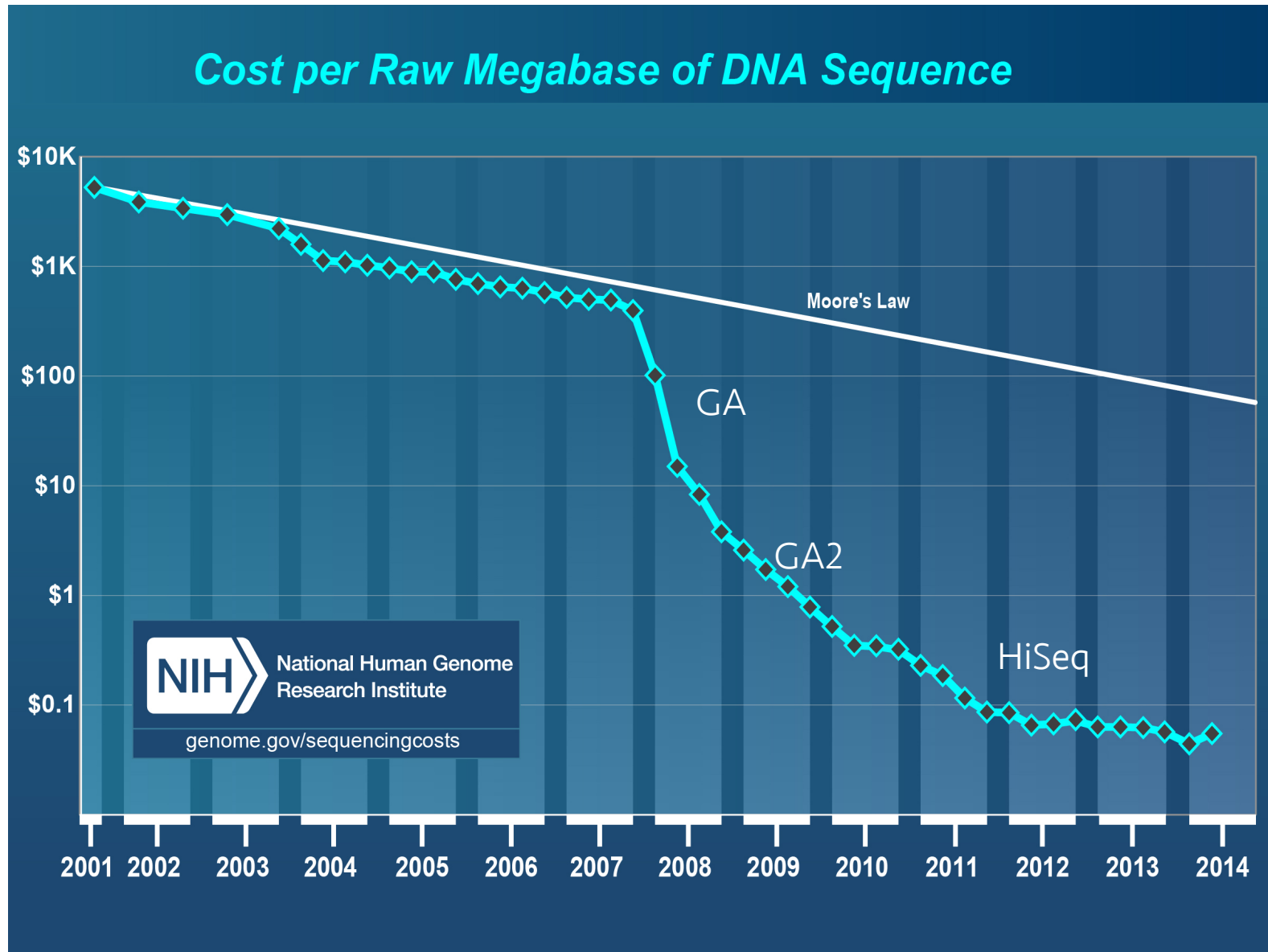
Single molecule sequencing
100 kB read lengths
15% error rate
Can detect DNA modifications



Which system to use?

- Depends on the application
 - Counting experiments (RNA/ChIP-Seq)
 - Genome assembly
 - Structural rearrangements
- Read numbers and length
 - Numbers– Illumina, Ion Torrent
 - Length – Pac Bio, Oxford Nanopore
- Error rates, types, bias
 - Illumina → substitution
 - Ion Torrent → Indel
 - PacBio/Oxford → high error rates (10-15%)

Sequencing costs have dropped dramatically



Instrument throughput has increased exponentially

Platform	Bases/read	Reads/run	Bases/run	Run/day	Bases/day	cost/MB
Sanger	700	96	6.7×10^5	24	1.6×10^6	\$500
Illumina GAI	150	10^8	1.5×10^{10}	1/8	1.8×10^9	\$0.520
Illumina HiSeq	250	4×10^9	1×10^{12}	1/6	1.6×10^{11}	\$0.029
Illumina XTen	300	6×10^9	1.8×10^{12}	1/3	6×10^{11}	\$0.007



You can sequence anything

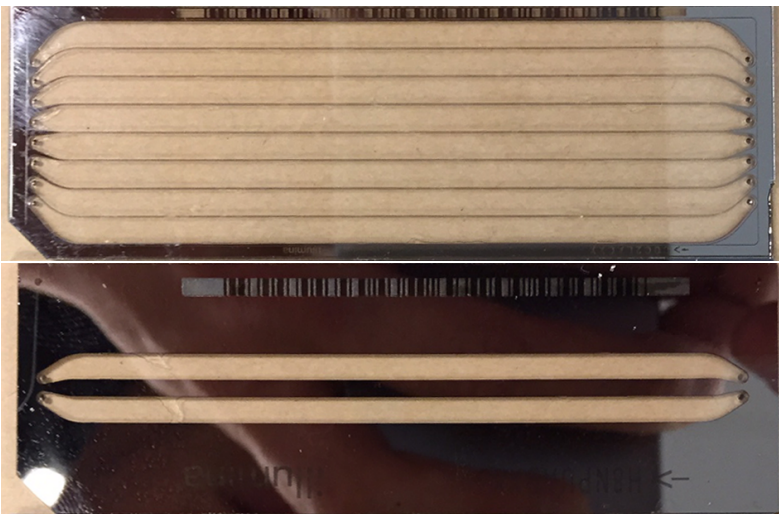
- Well, almost
 - Make DNA
 - Add adapters
 - Total size <1kb
- Many ways to add adapters
 - PCR
 - Ligation
 - Reverse transcription
 - Primer extension

You can sequence anything

- Counting applications
 - RNA-Seq
 - CHIP-Seq
 - Footprinting (ribosome, transcription,...)
- Genome
 - Whole-genome (was \$1000s/genome)
 - Exome – pull out coding sequence
 - DNA methylation (bisulfite conversion)
 - metagenomics

Sequencing is really, really cheap

- \$1000 human genome
- Non-invasive prenatal testing
- Large population studies
 - Pick out SNPs and mutations responsible for disease
 - Tumor/normal sequencing
 - Whole-genome sequencing will be universal
- Mostly due to Illumina (but watch out for others)



Flow cells, where the magic happens

Illumina library

Insert

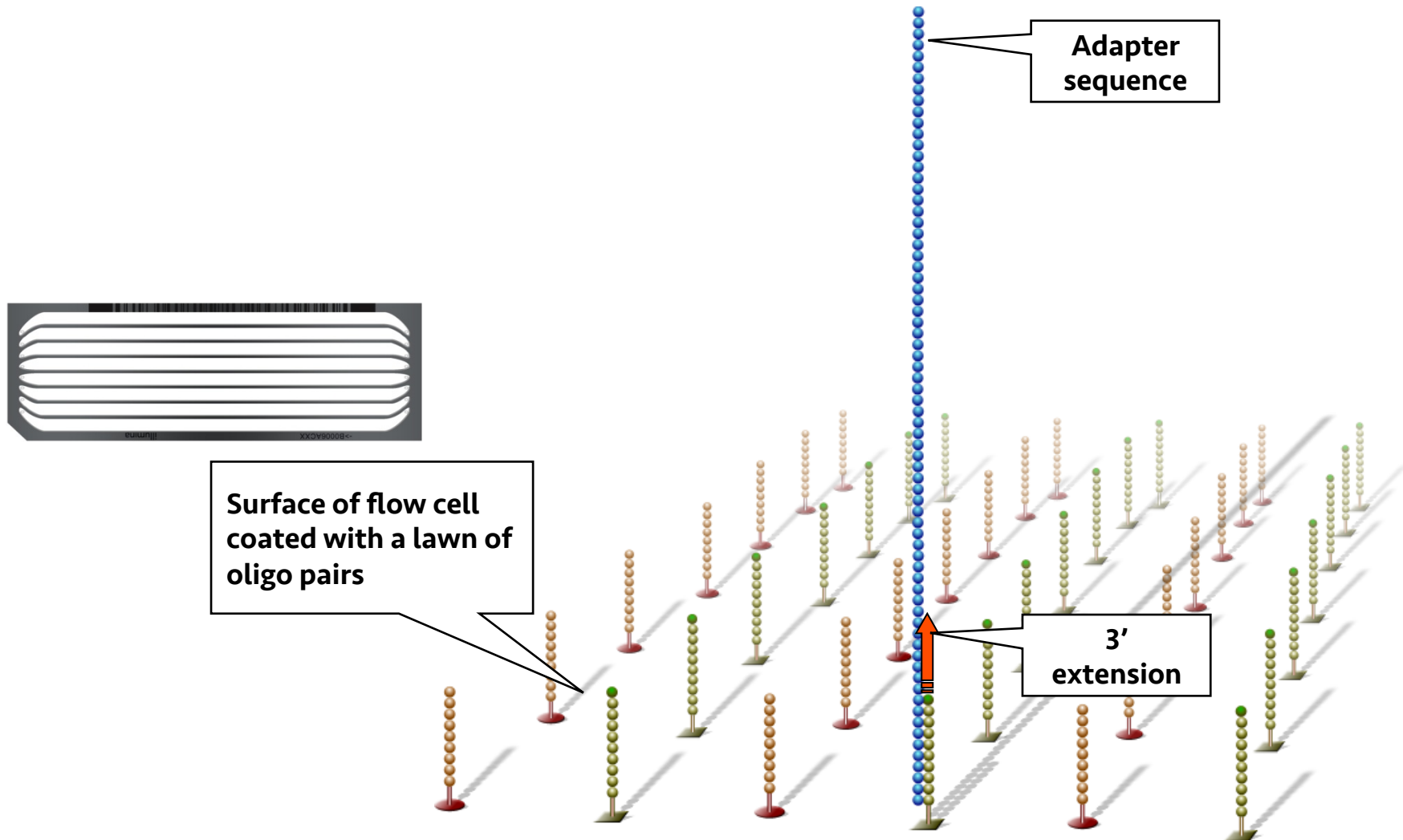


Illumina library

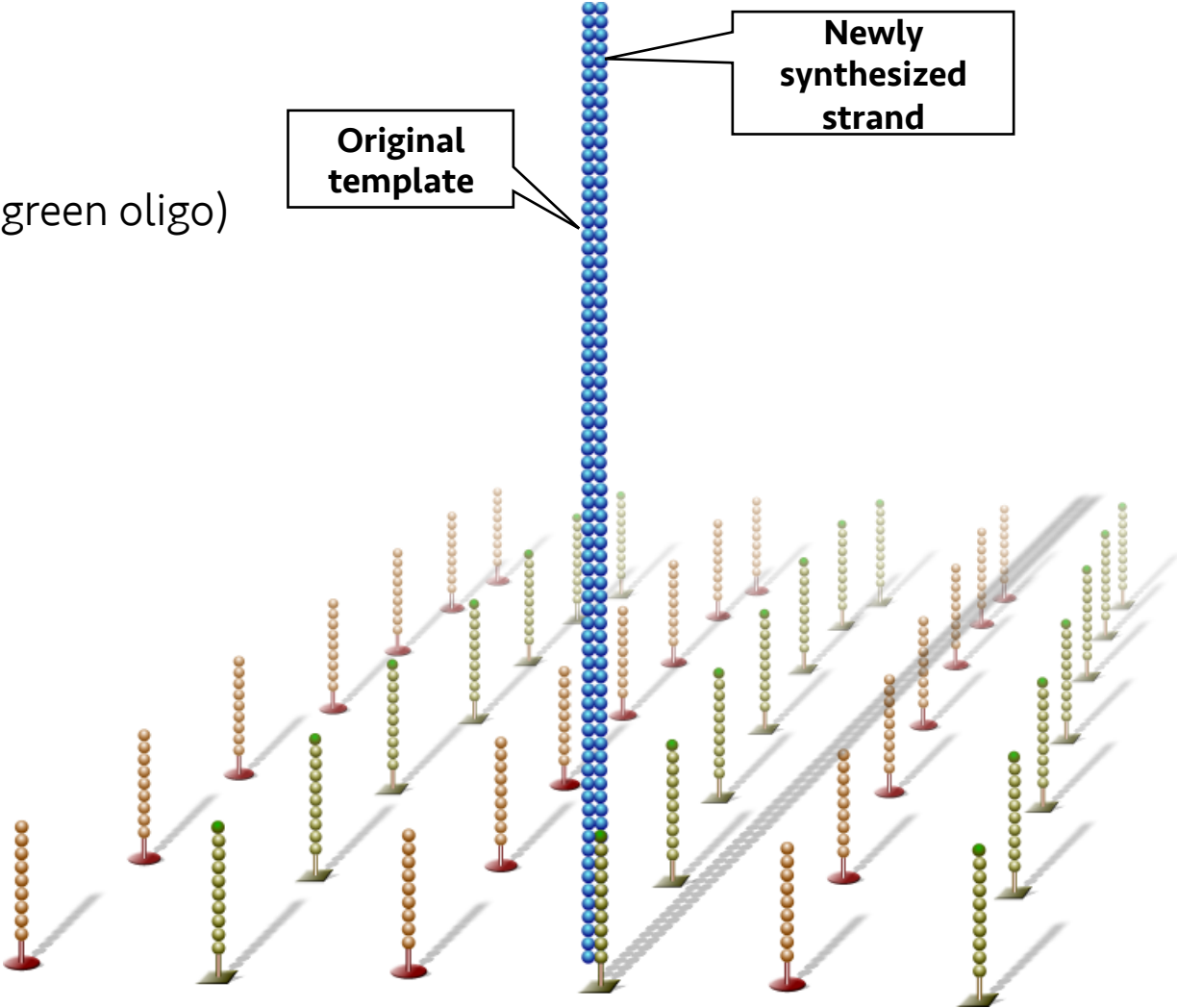


- Adapter sequence includes primer binding sites and capture sequences.
- Ion Torrent library adapters are very similar.

Library binding to flow cell

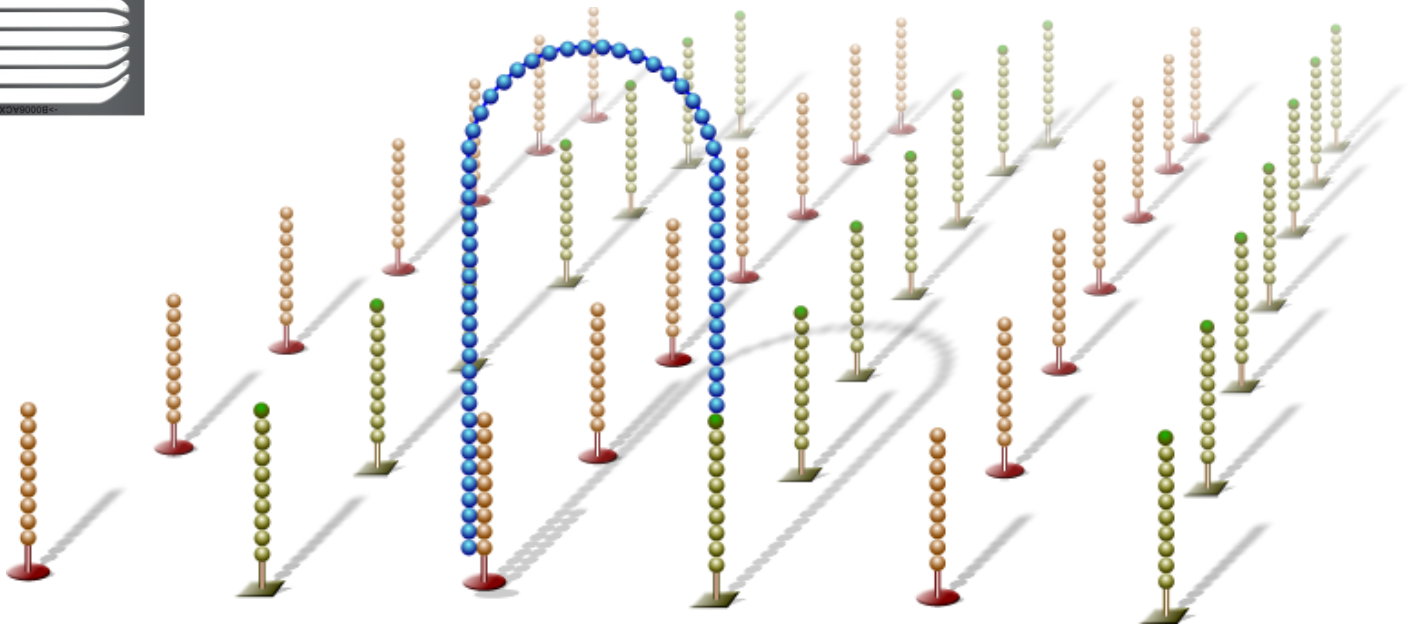


Reverse strand is created (green oligo)

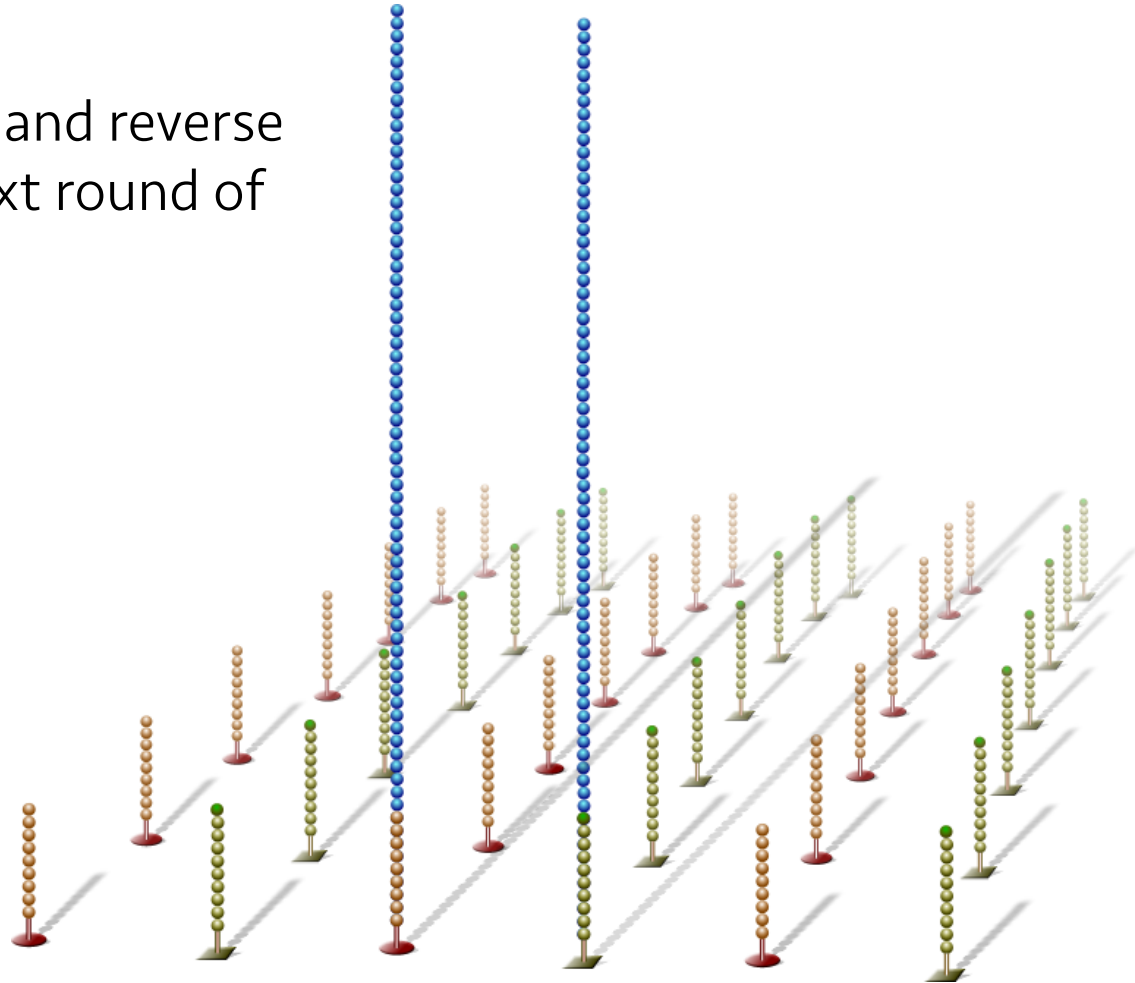


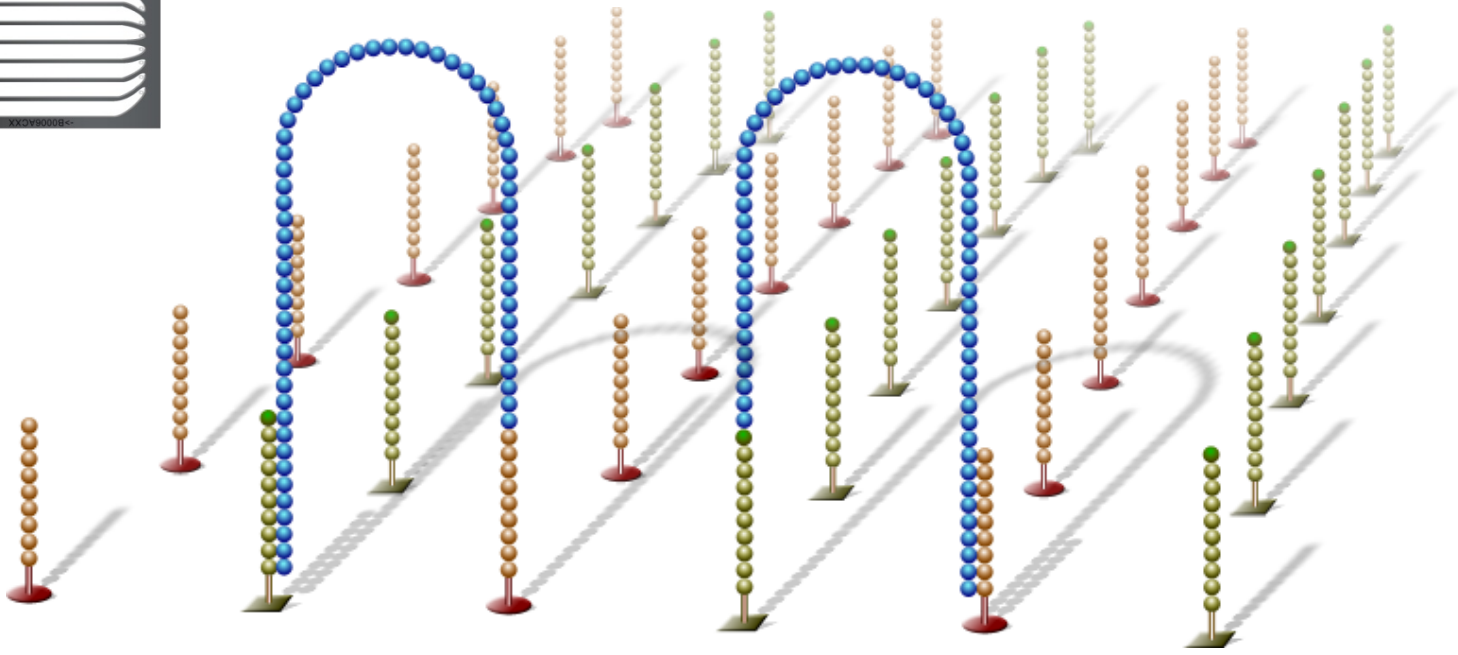
Original template

Newly synthesized strand

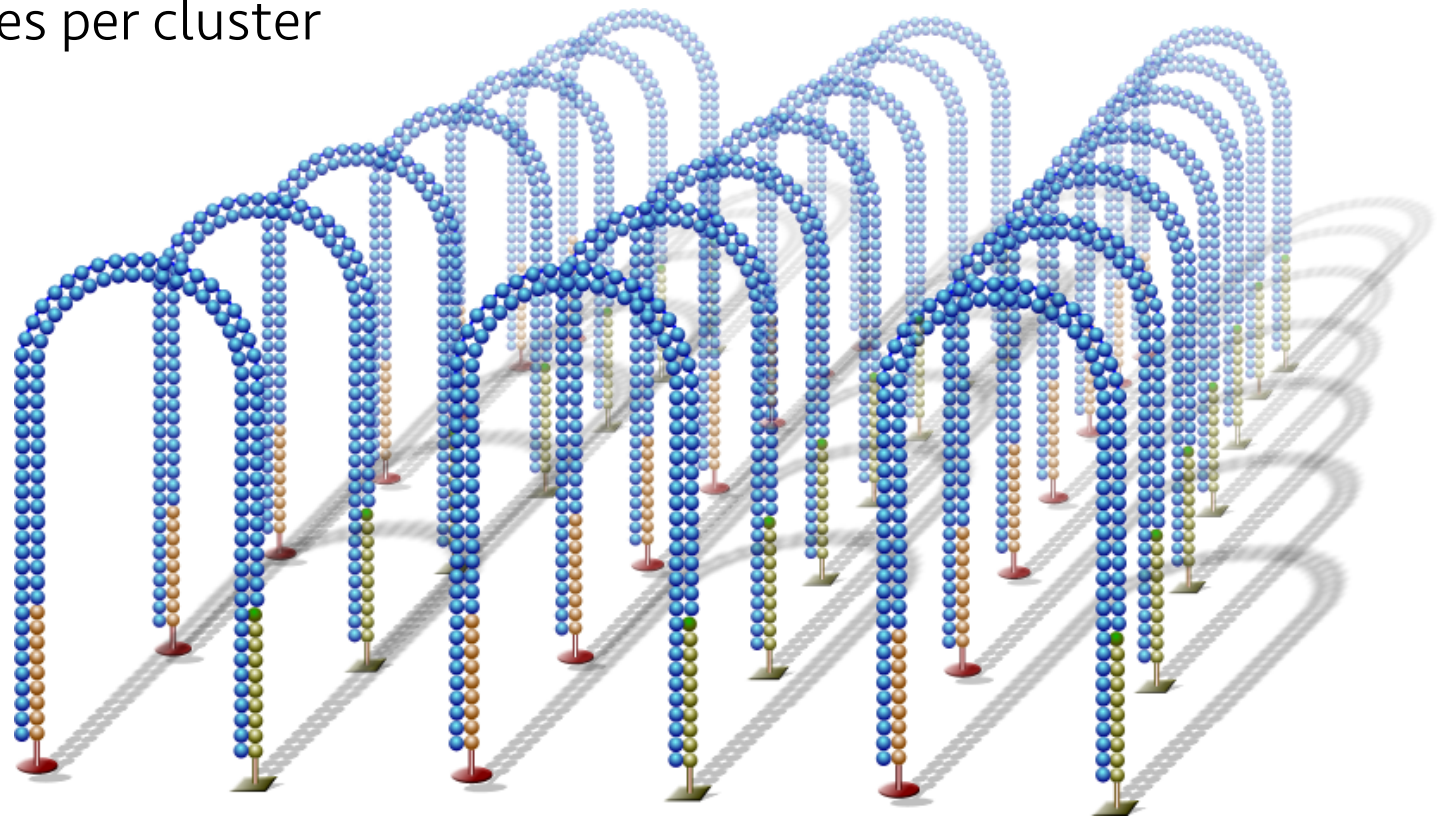


Original (orange oligo) and reverse strand are ready for next round of PCR .

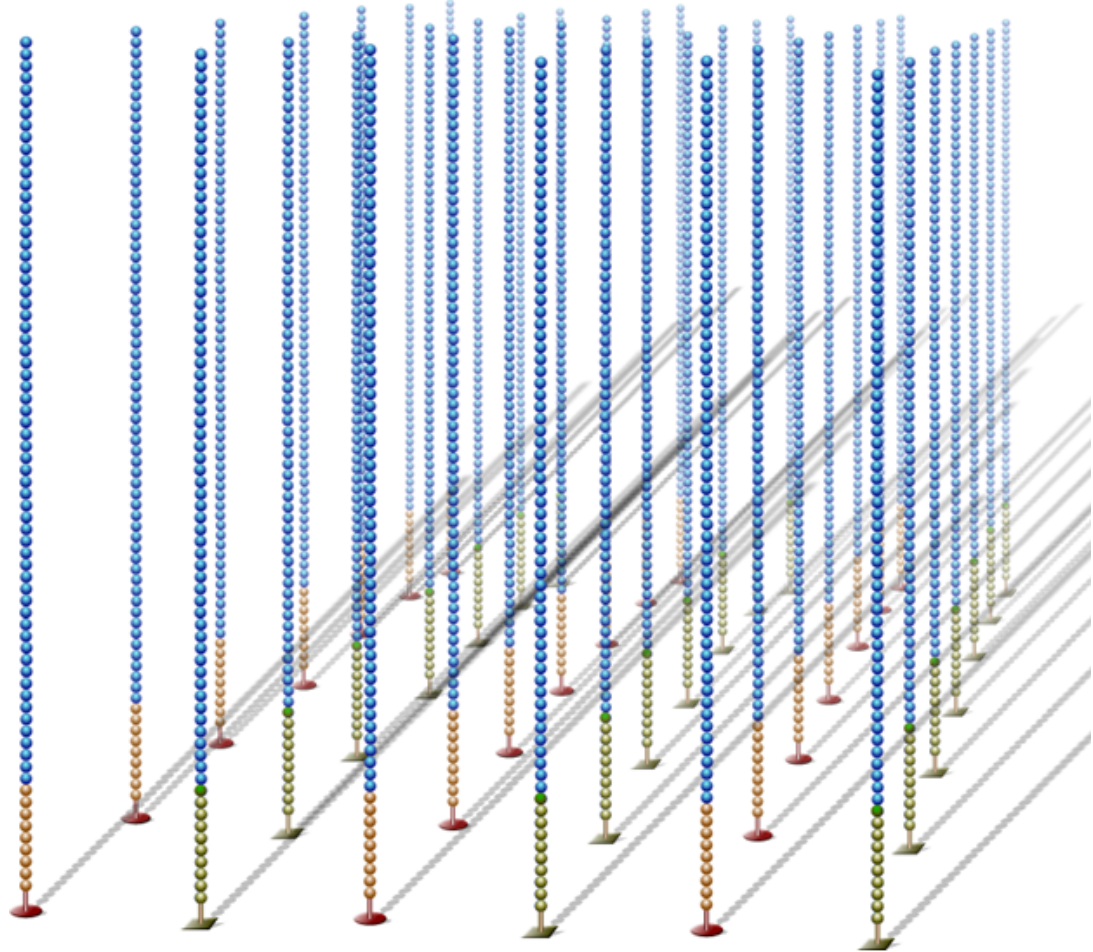




At the end of clustering
~1000 copies per cluster

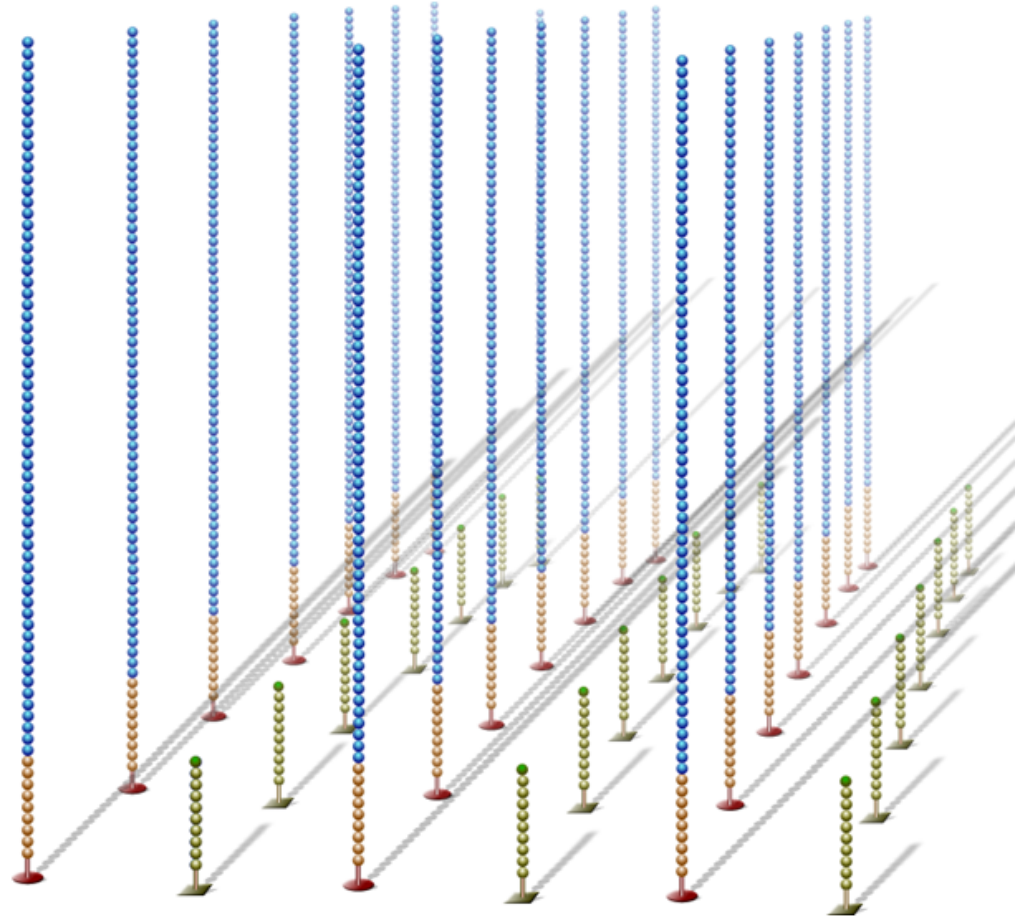
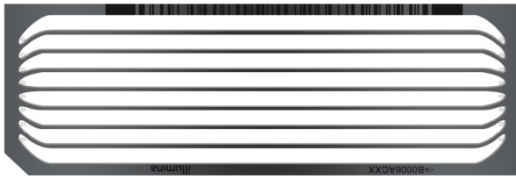


Molecules are linearized

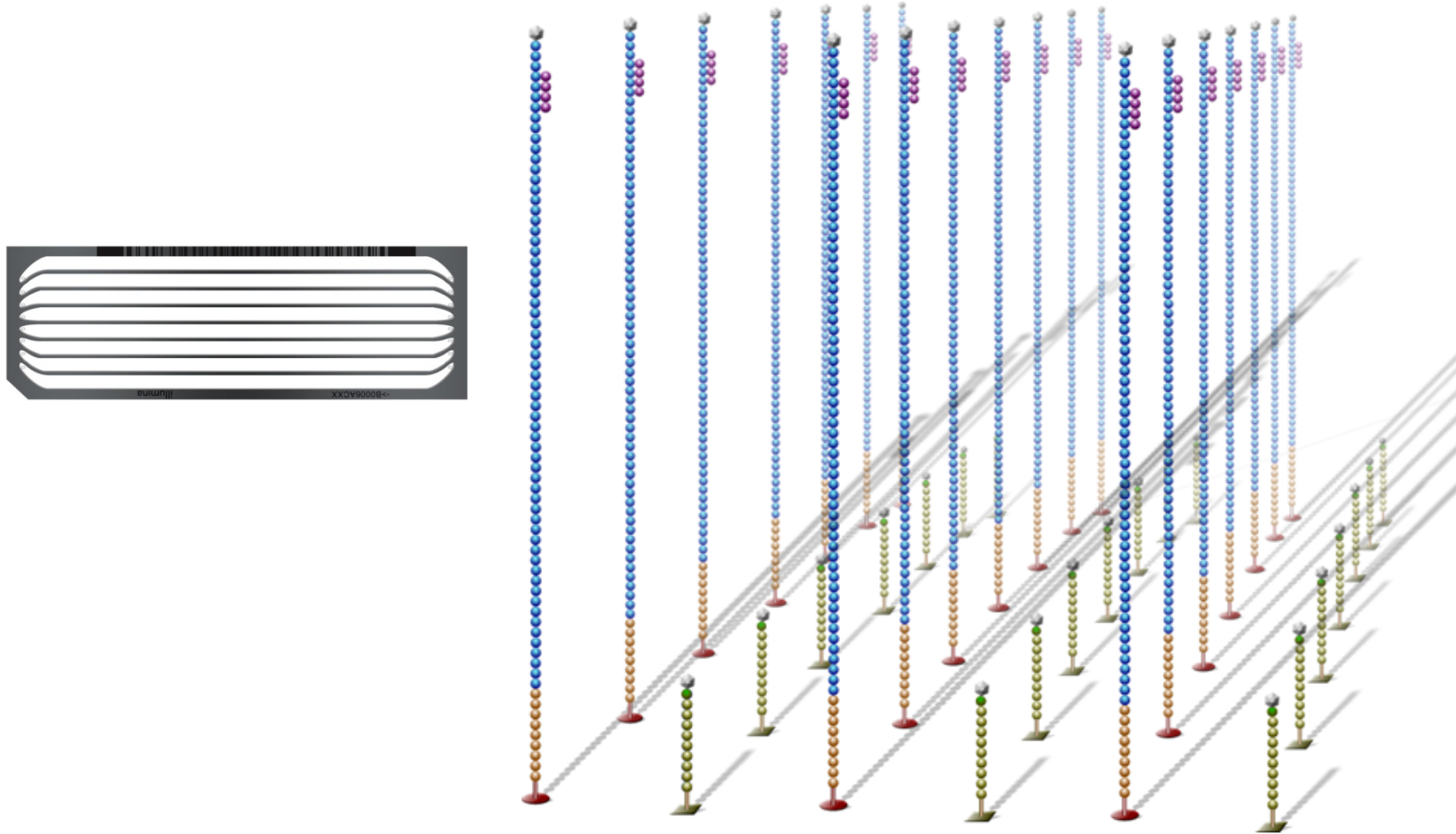


Reverse Strands are cleaved

Original template strand remains (orange oligo)



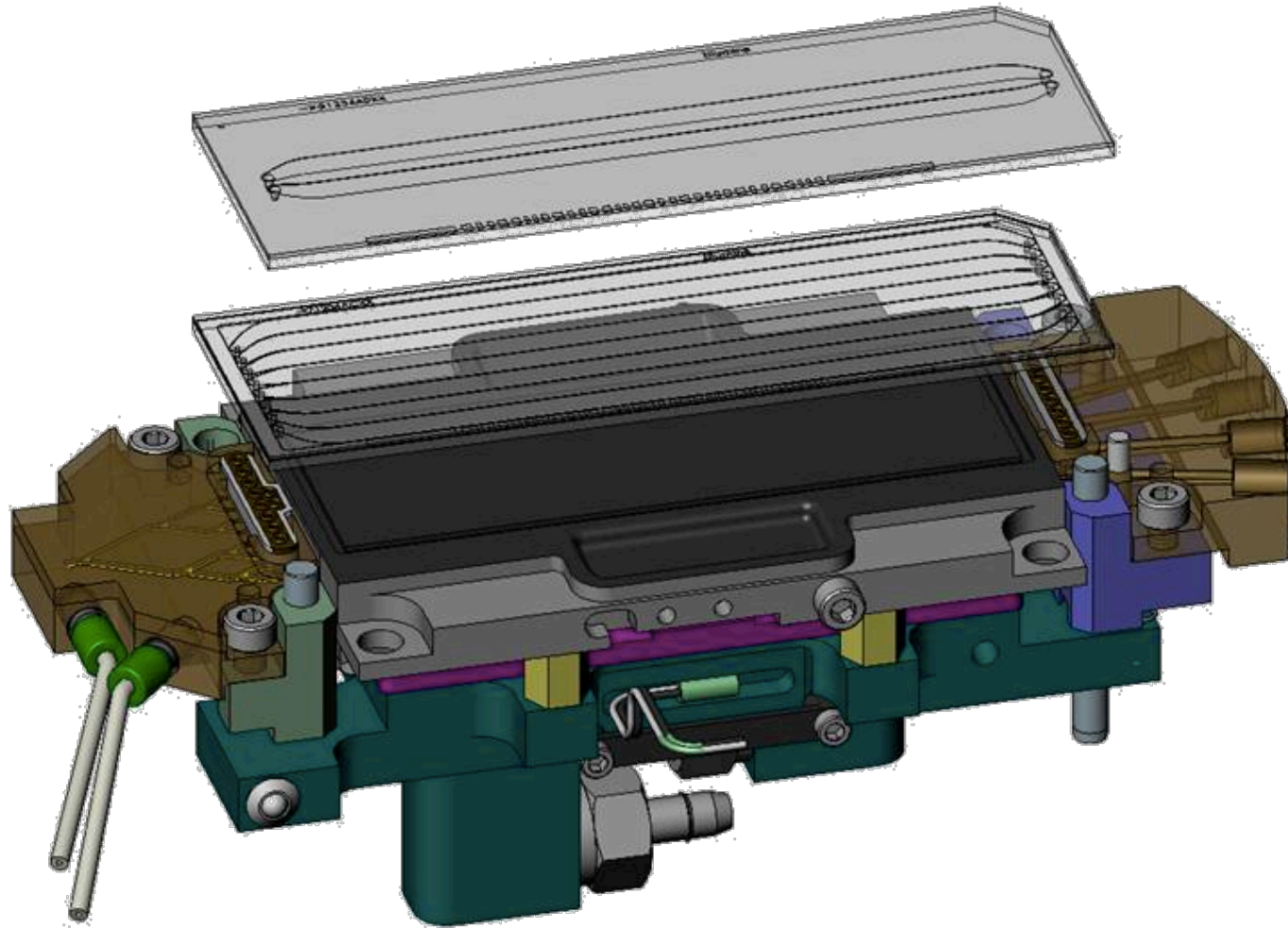
Ends are blocked and sequencing primers hybridized



After clustering flow cell moves to the HiSeq

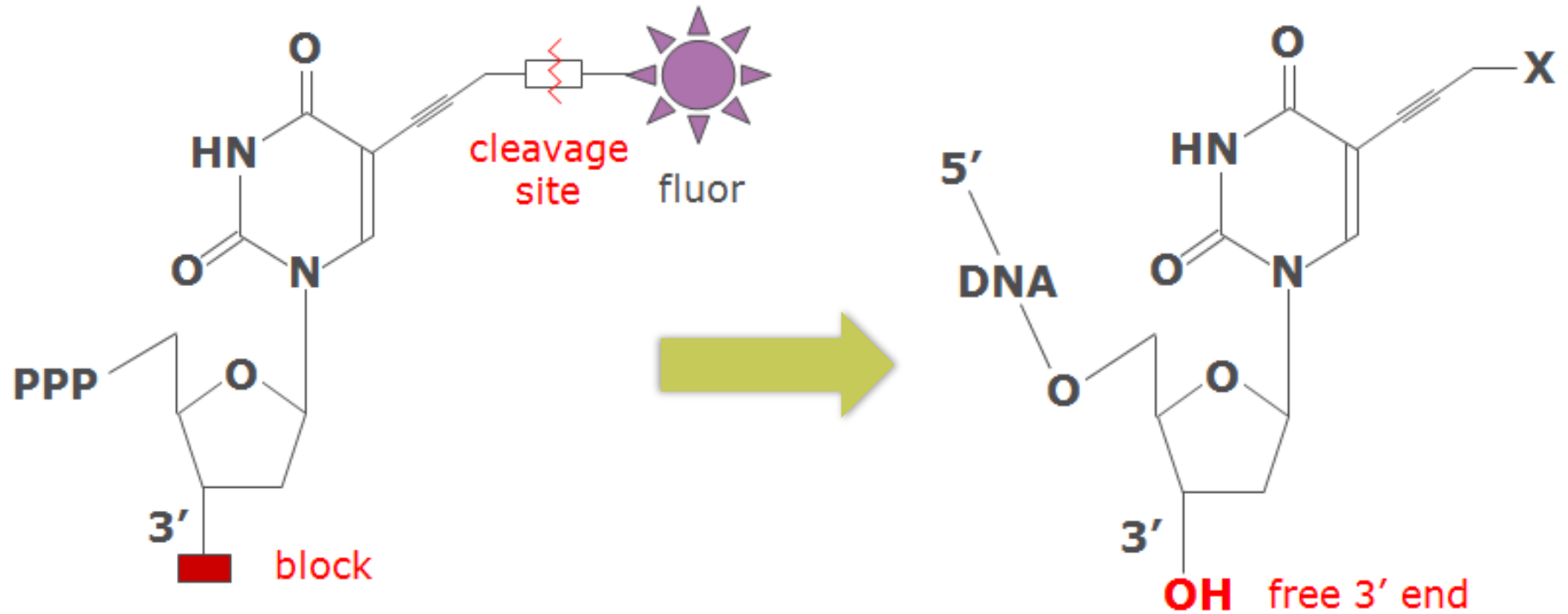


Sequencer is a microscope with fluidic channels

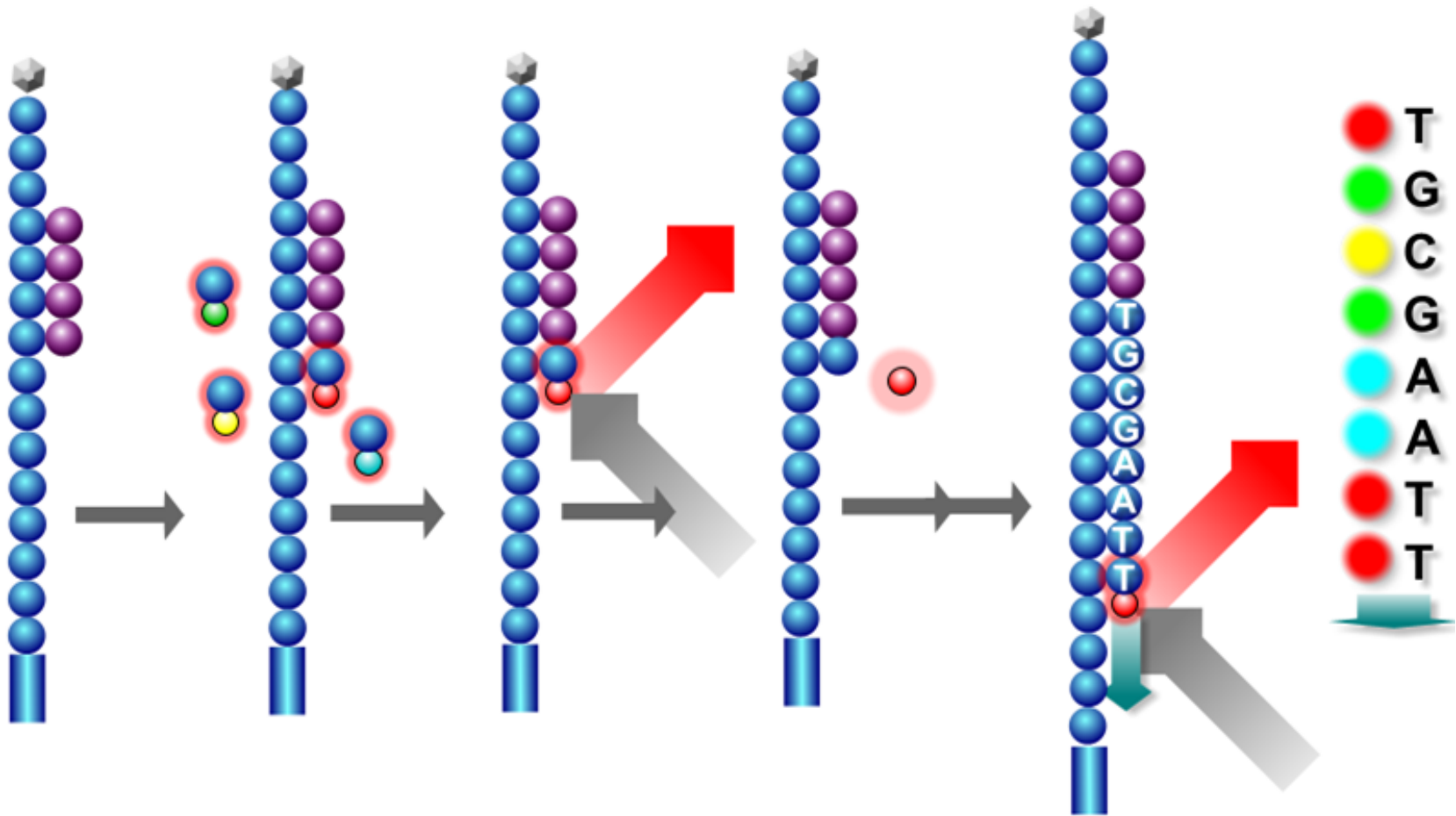


Reversible Terminator Chemistry

- A
- C
- G
- T



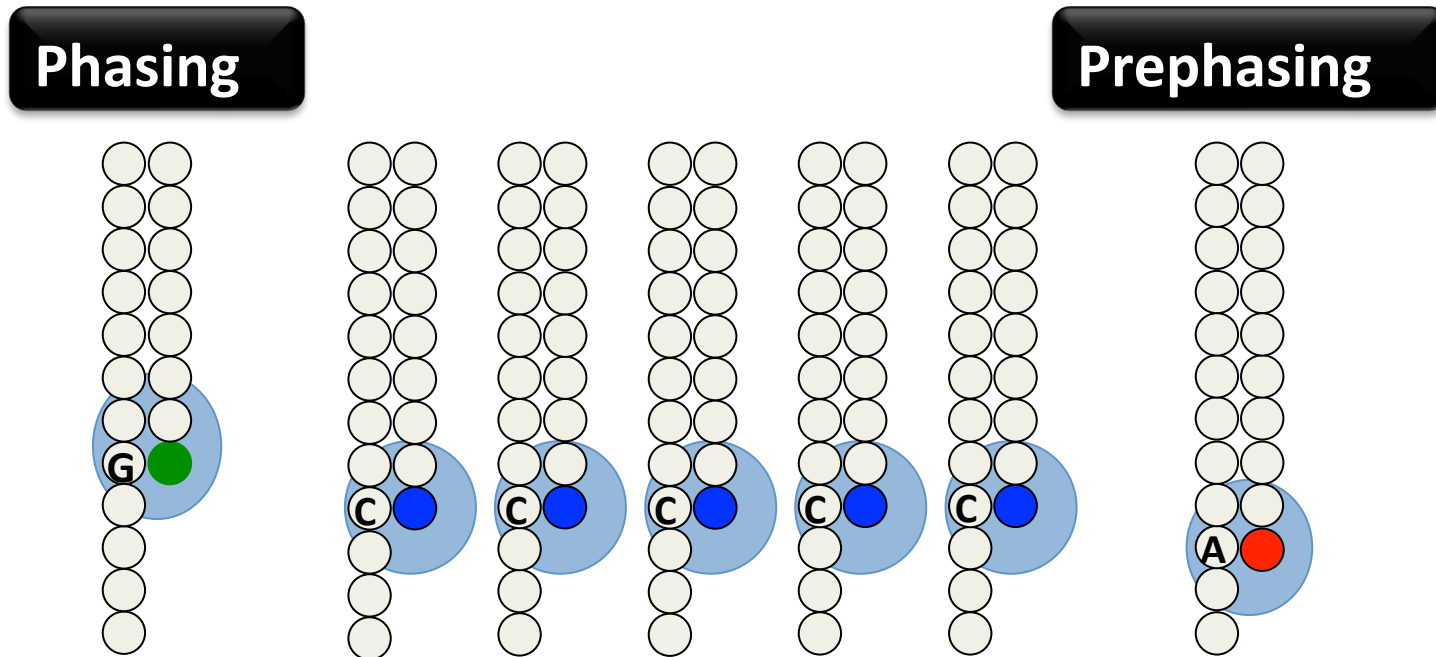
Illumina SBS technology



Limitations of SBS

Each clonal cluster contains ~1,000 copies

Imperfect chemistry → some strands will lag and others will jump ahead



This limits the length of runs.

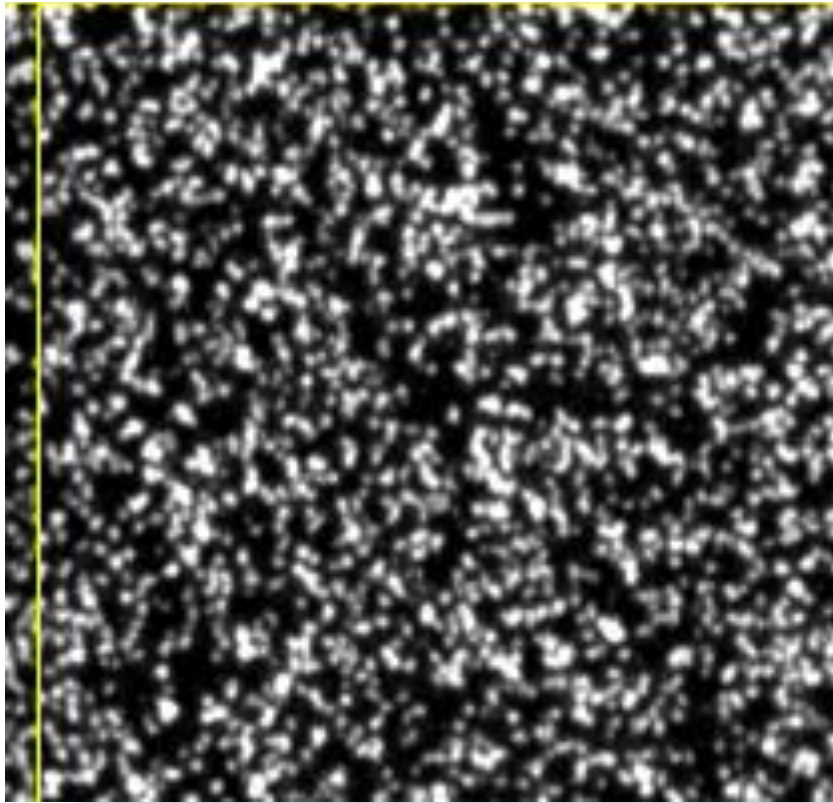
HiSeq 2x150

MiSeq 2x300

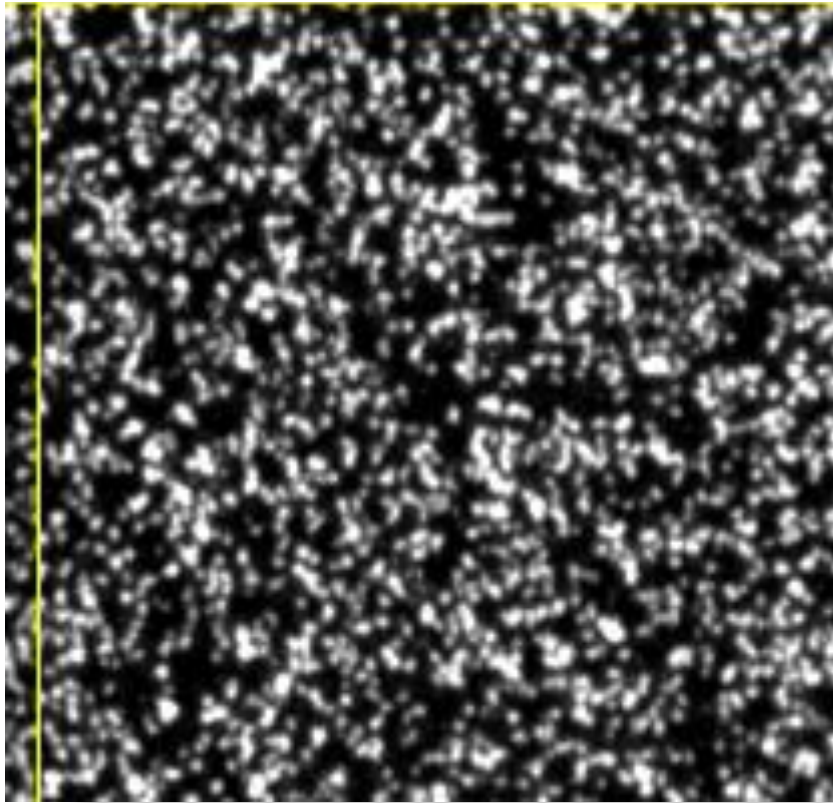
Going from images to sequence

- Find clusters
- Calculate intensities
- Make basecalls

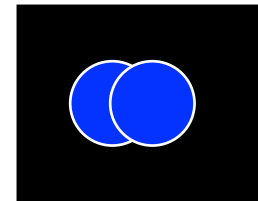
Sequence diversity is critical for template generation



Sequence diversity is critical for template generation

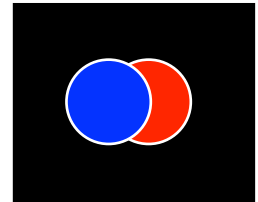


CYCLE 1



G channel

CYCLE 2

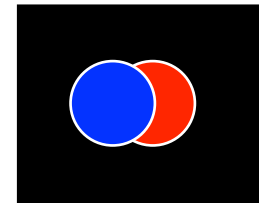


C channel

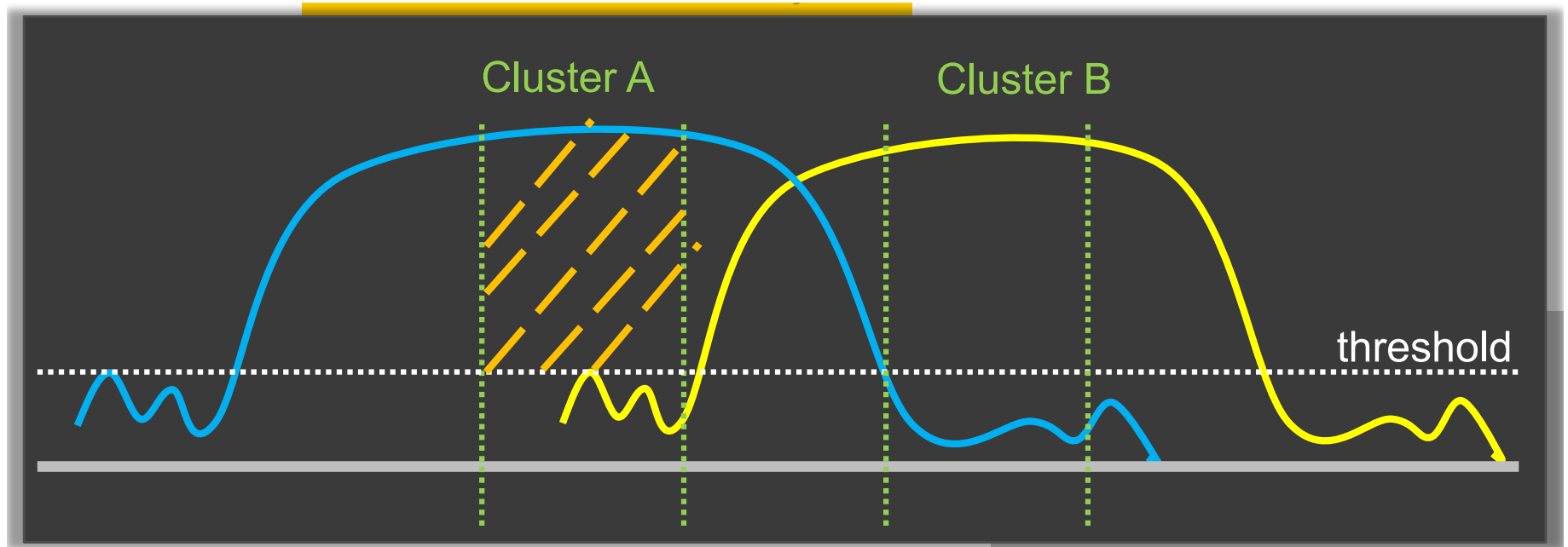
G channel

Sequence diversity is critical for template generation

CYCLE 2

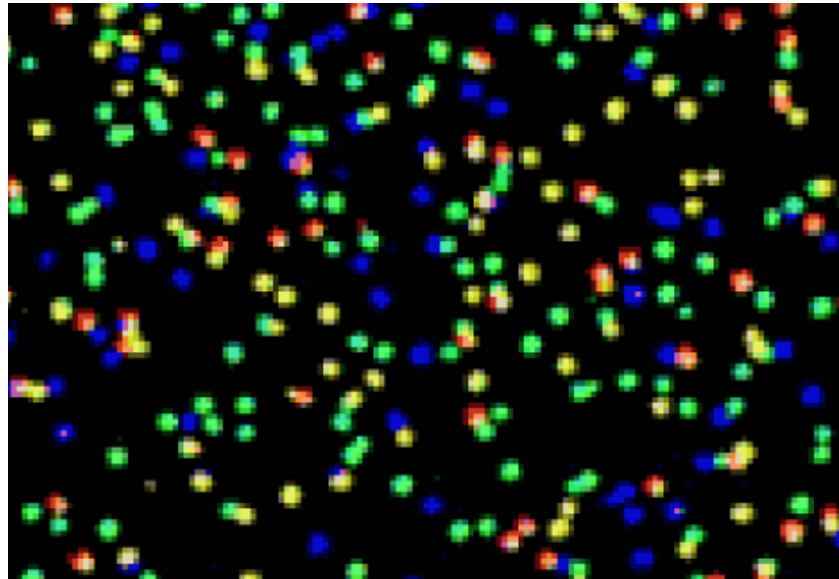


C channel
G channel



Raw images

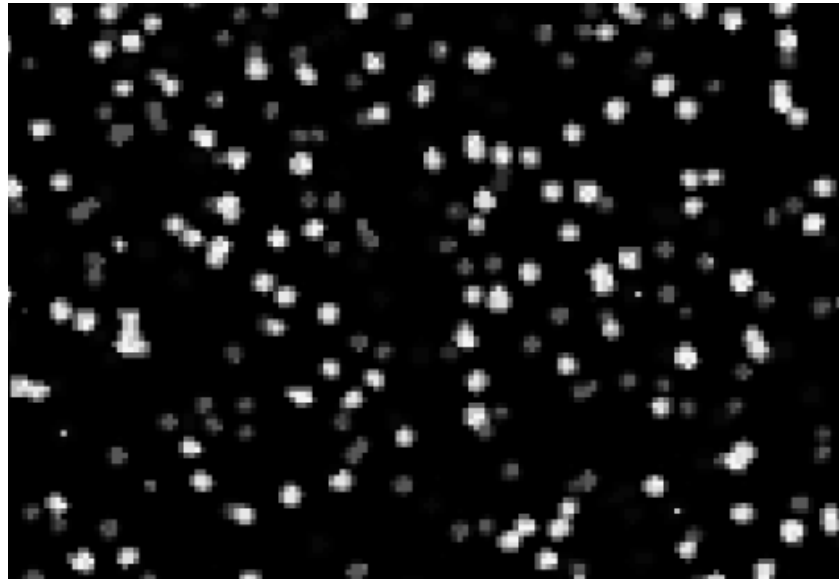
System takes 4 images each cycle, 1 per nucleotide



Raw images

System takes 4 images each cycle, 1 per nucleotide

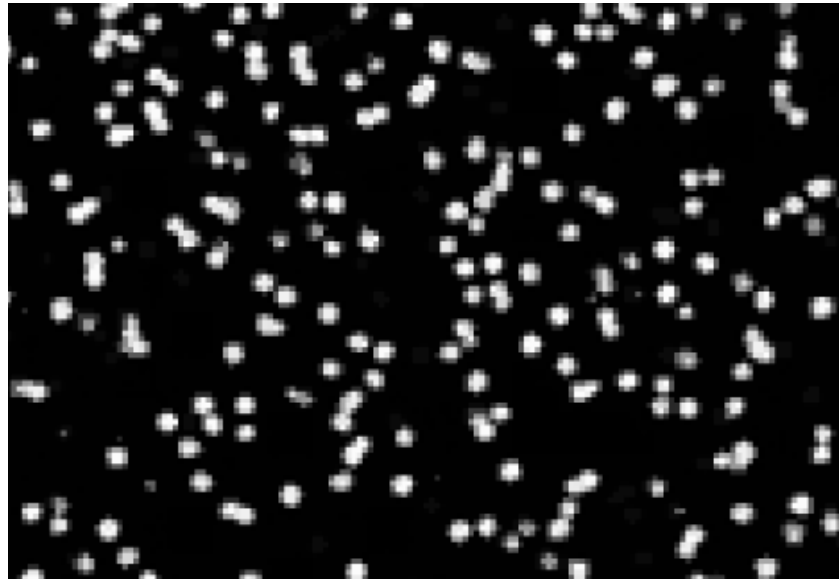
Red



Raw images

System takes 4 images each cycle, 1 per nucleotide

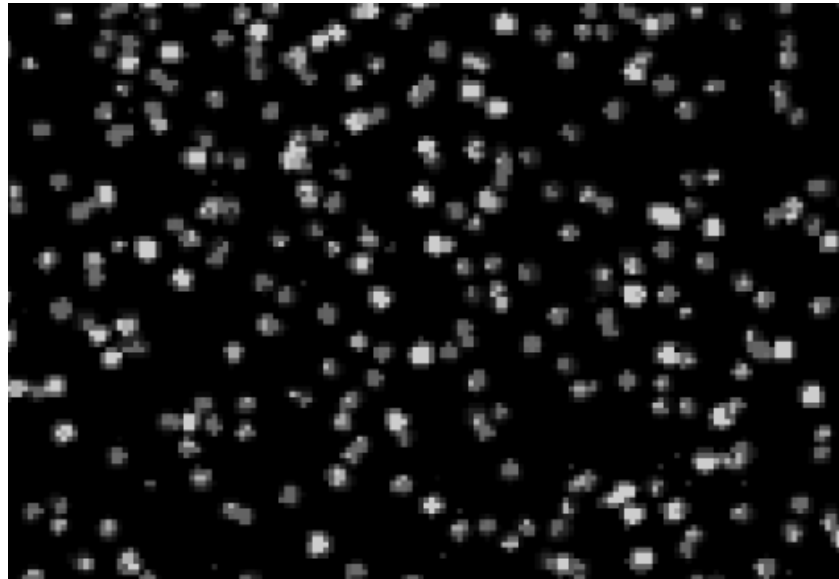
Green



Raw images

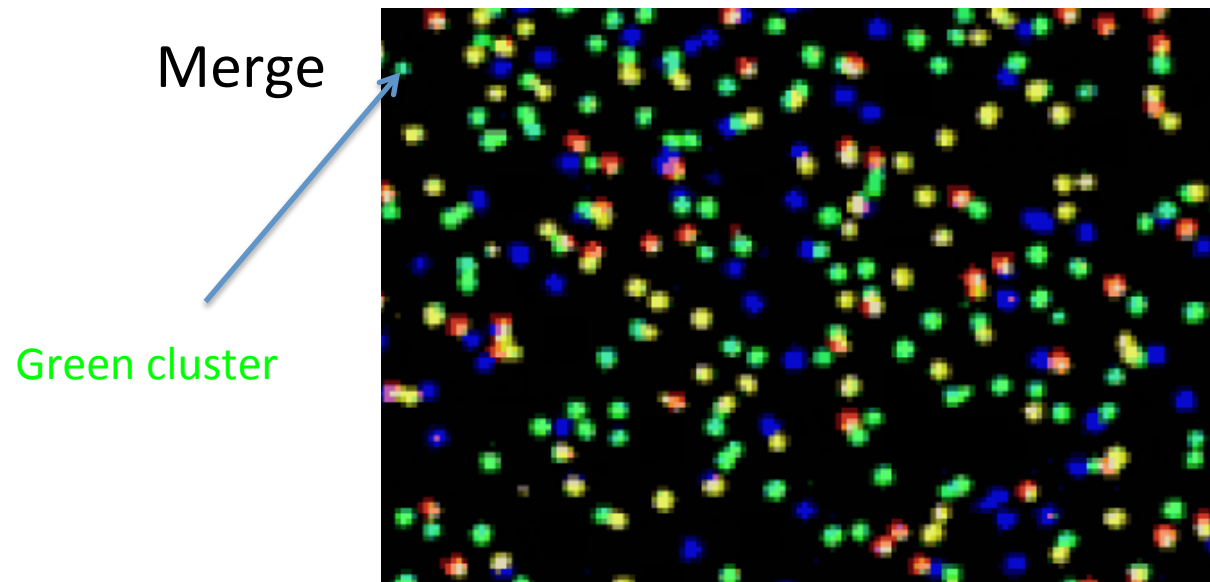
System takes 4 images each cycle, 1 per nucleotide

Blue



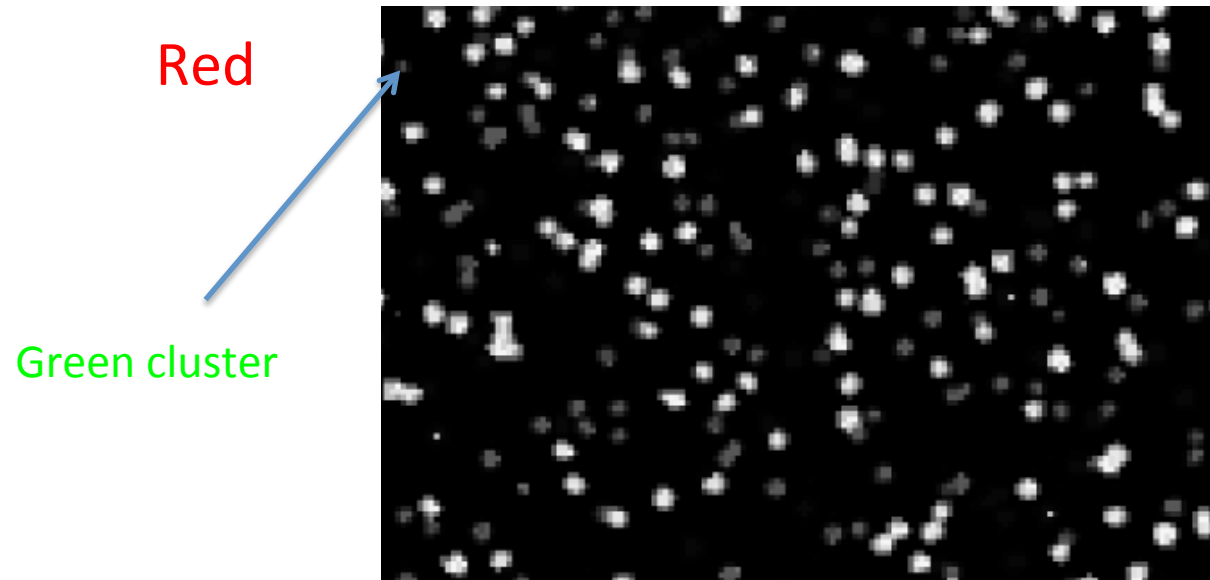
Raw images

System takes 4 images each cycle, 1 per nucleotide



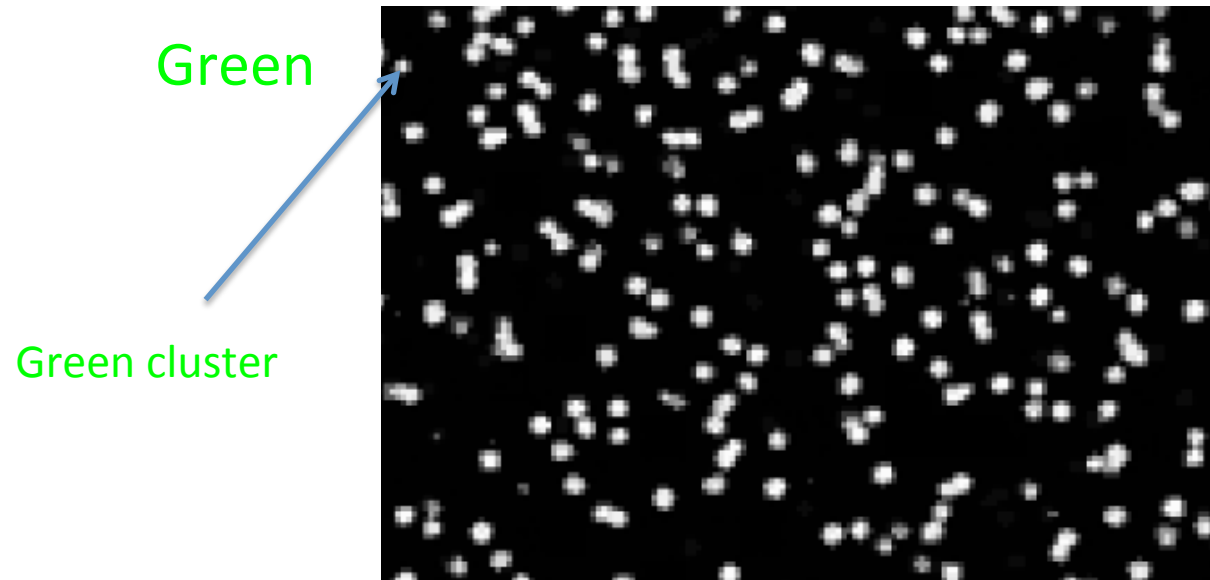
Fluorescence crosstalk

Fluorescent label spectra overlap



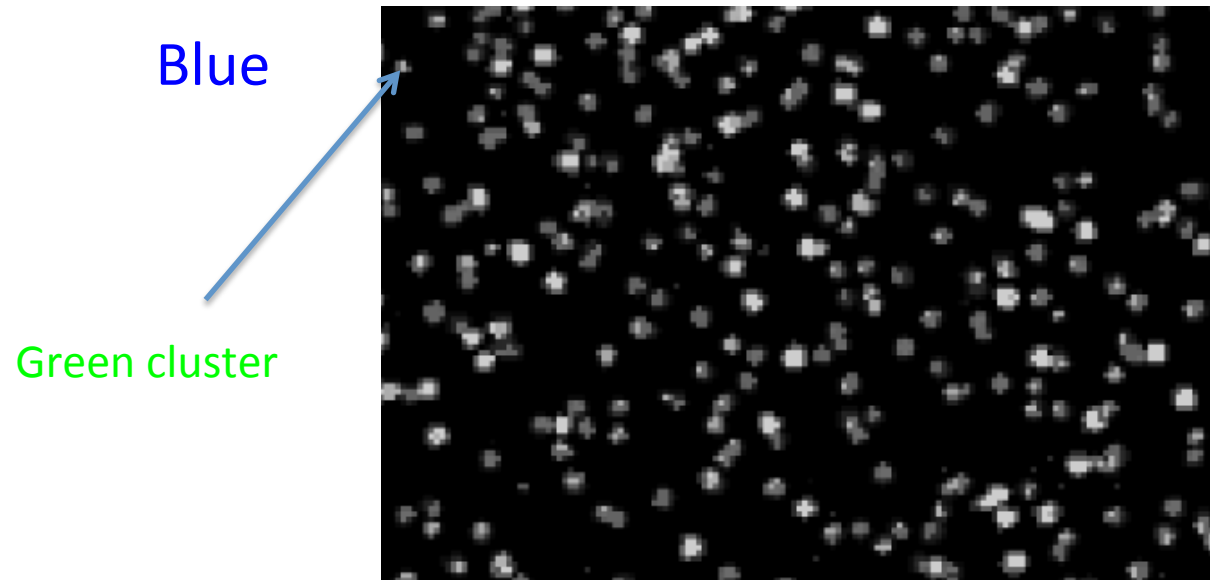
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Fluorescent label spectra overlap

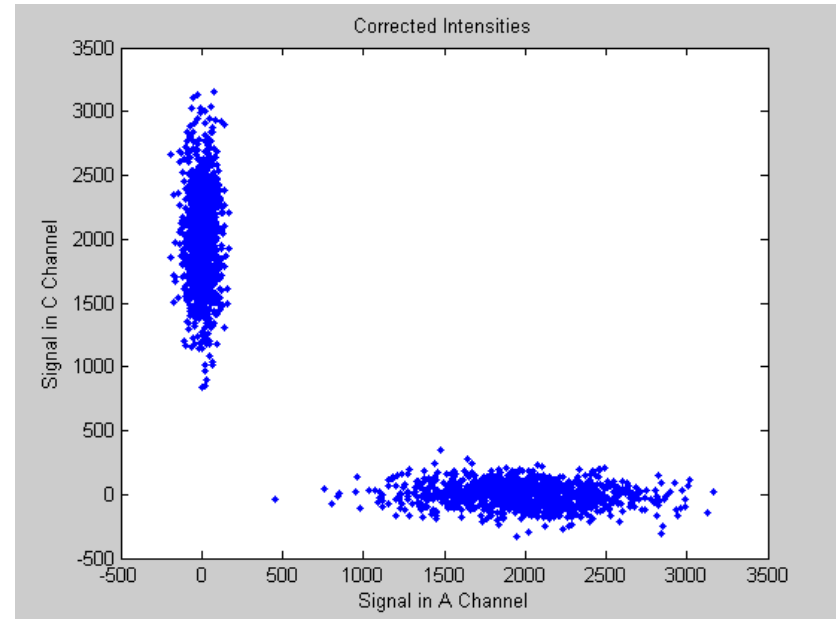
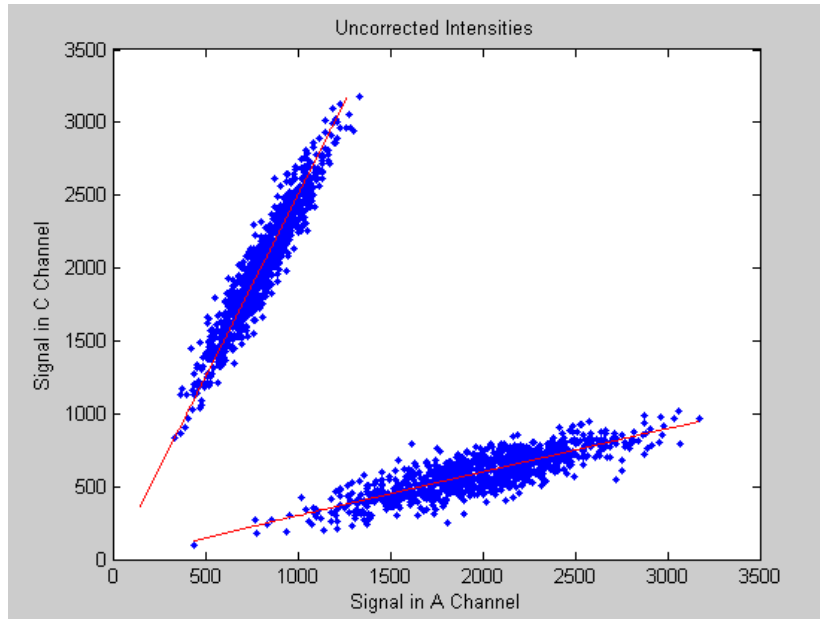


Fluorescence crosstalk

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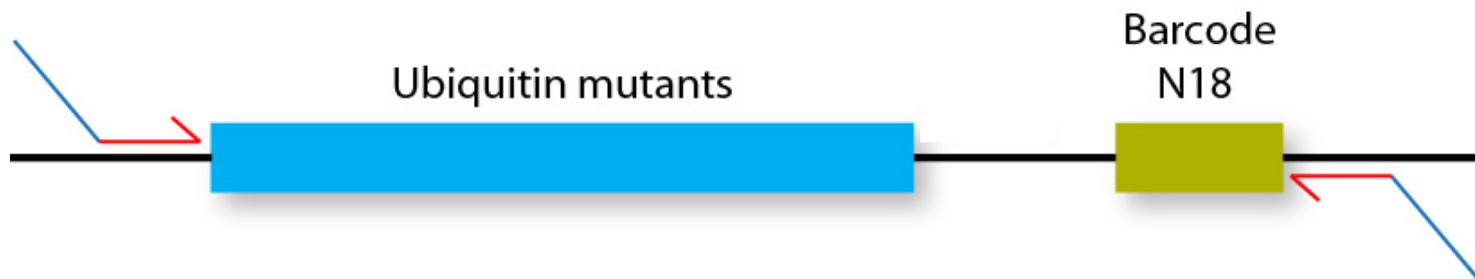
Signal cross talk correction



Ubiquitin library



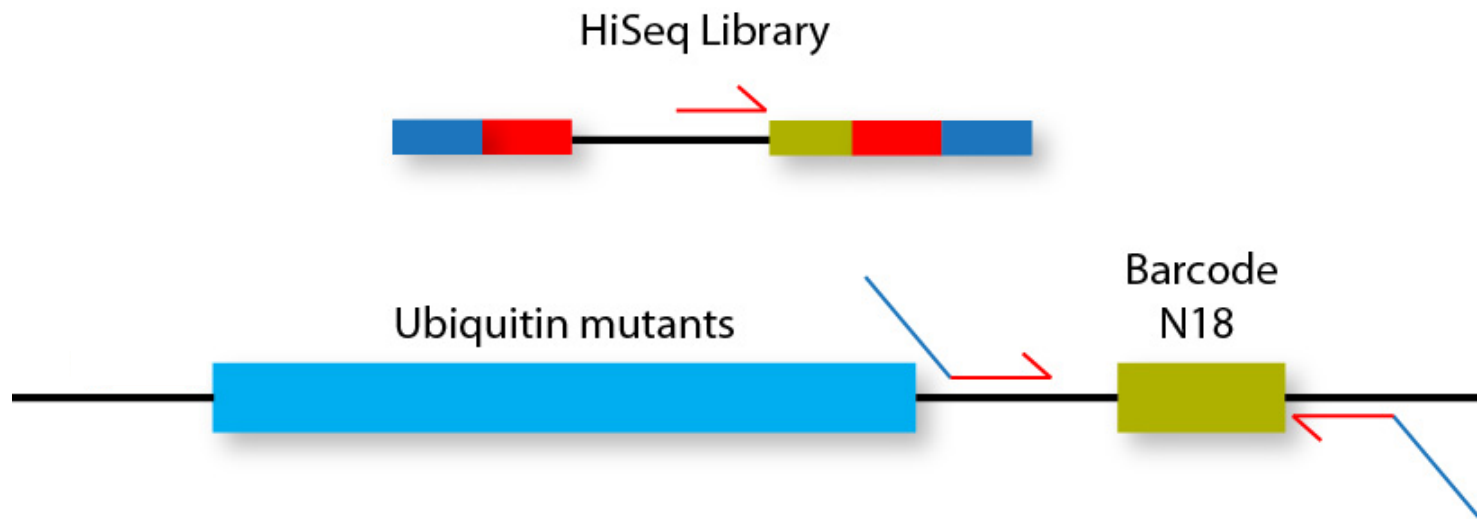
Ubiquitin library



MiSeq Library - link barcode with ubiquitin mutant



Ubiquitin library



- HiSeq is much cheaper than the MiSeq
 - 10-20x more reads
 - 20% more expensive
 - Save MiSeq for longer reads